

# Measurements, applications, and impact of the refractive index of extracellular vesicles

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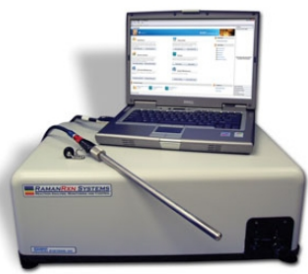
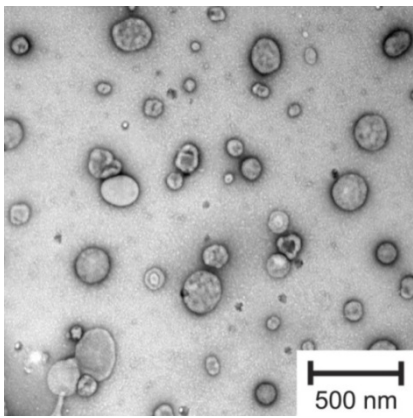
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Presentation includes discussion of the following off-label use of a drug or medical device:

<N/A>

# Vesicle refractive index is essential



# Timeline vesicle detection with optical methods

Flow cytometry standardization <i>Lacroix et al.</i>	Flow cytometry calibration <i>Chandler et al.</i>	Scanning flow cytometry <i>Konokhova et al.</i>	Flow cytometry artifact <i>Nolan et al.</i>	Flow cytometry standardization by vesicle size approximation <i>Coumans et al.</i>
	Nanoparticle tracking analysis <i>Dragovic et al.</i>	Raman on vesicles <i>Tatischeff et al.</i>		

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# Timeline vesicle detection with optical methods

Flow cytometry  
standardization  
*Lacroix et al.*

Flow cytometry  
calibration  
*Chandler et al.*

Scanning flow  
cytometry  
*Konokhova et al.*



Nanoparticle  
tracking analysis  
*Dragovic et al.*

Raman  
vesicles  
*Tatischeff et al.*

Flow cytometry  
standardization  
vesicle size  
approximation  
*Smans et al.*

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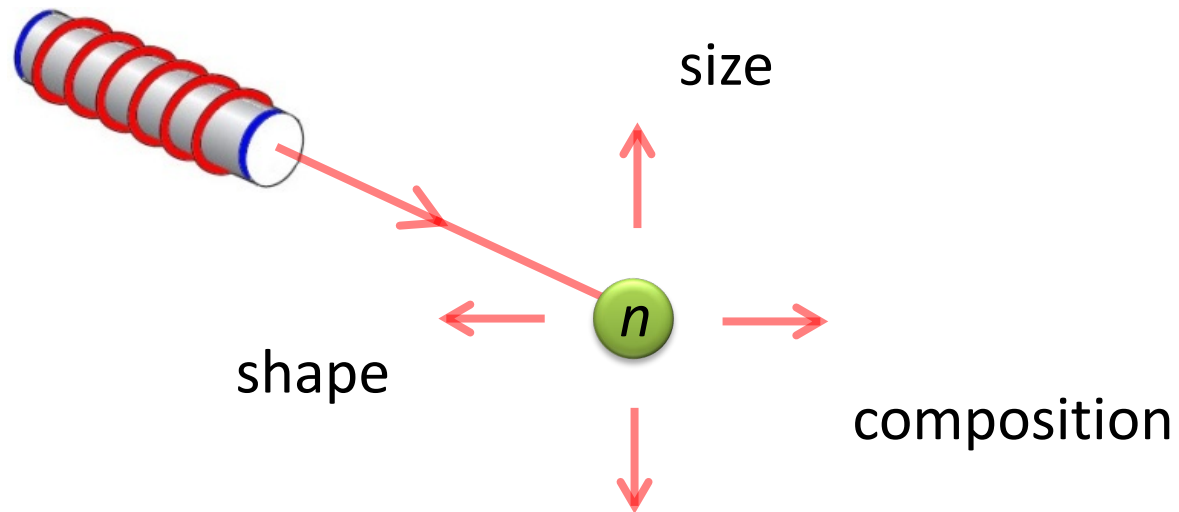
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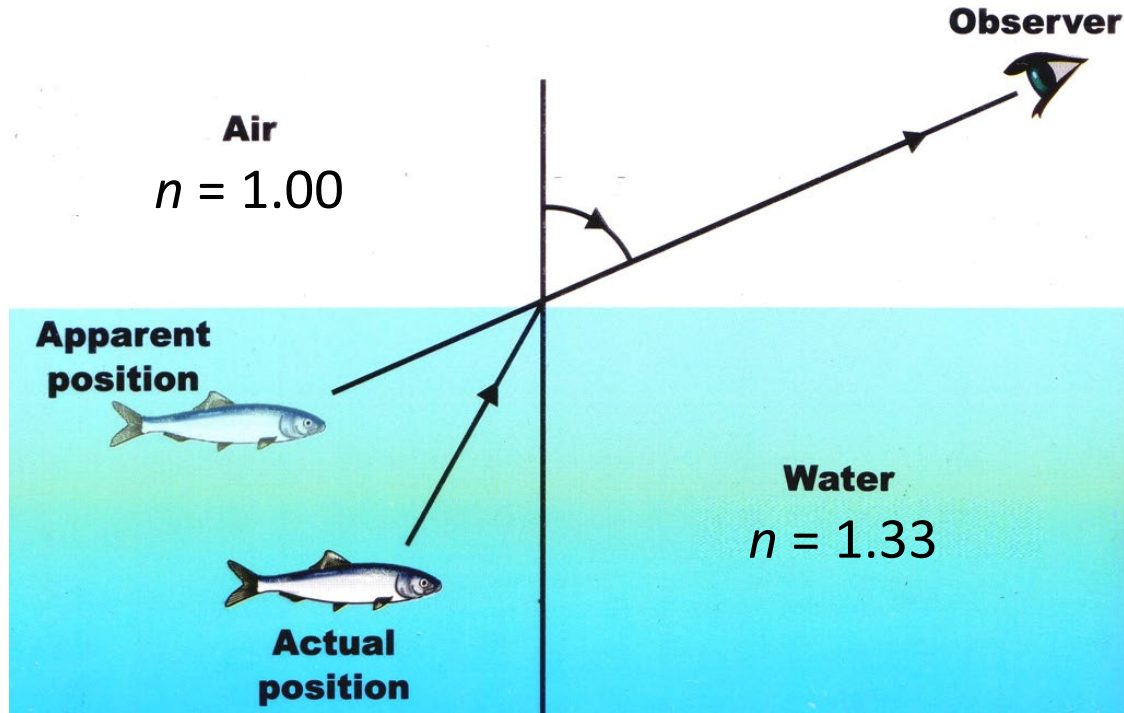
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# Introduction to light scattering



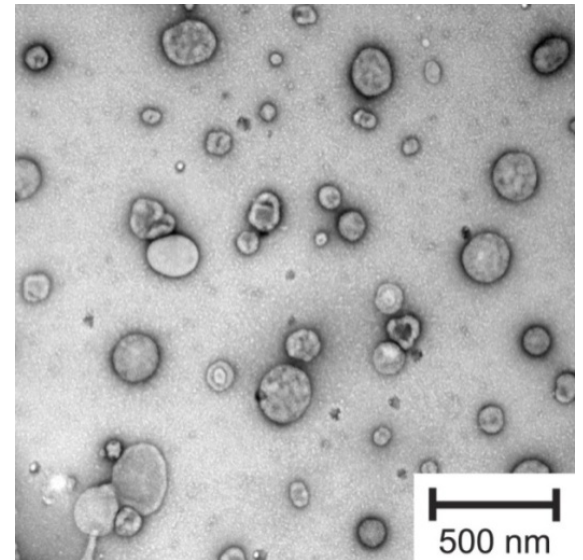
- light illuminating a vesicle is partly absorbed and partly scattered (deflected)
- light scattering depends on size and refractive index ( $n$ )

# Introduction to the refractive index ( $n$ )



# Problem

- no technique is capable of determining the refractive index of particles being
  - <500 nm
  - heterogeneous in size
  - heterogeneous in refractive index
  - in suspension



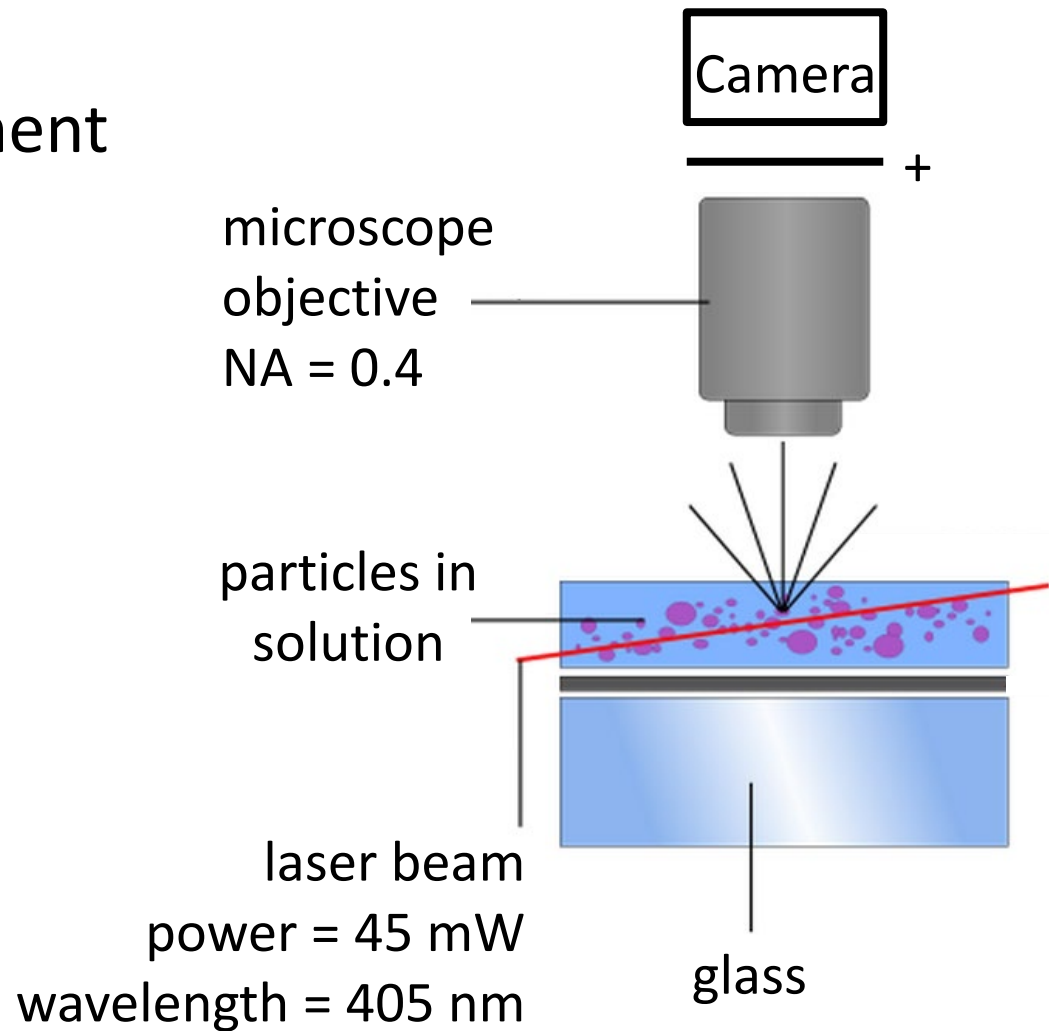


# Goal

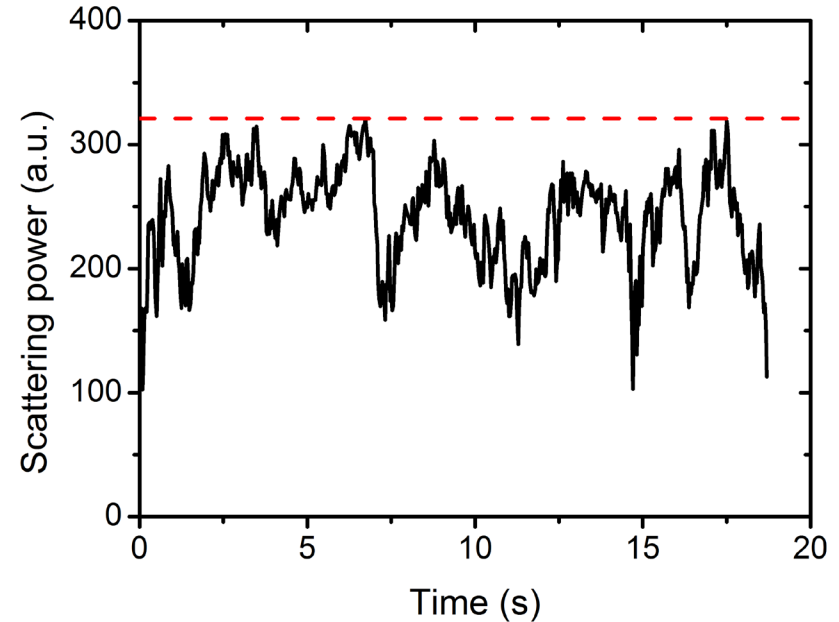
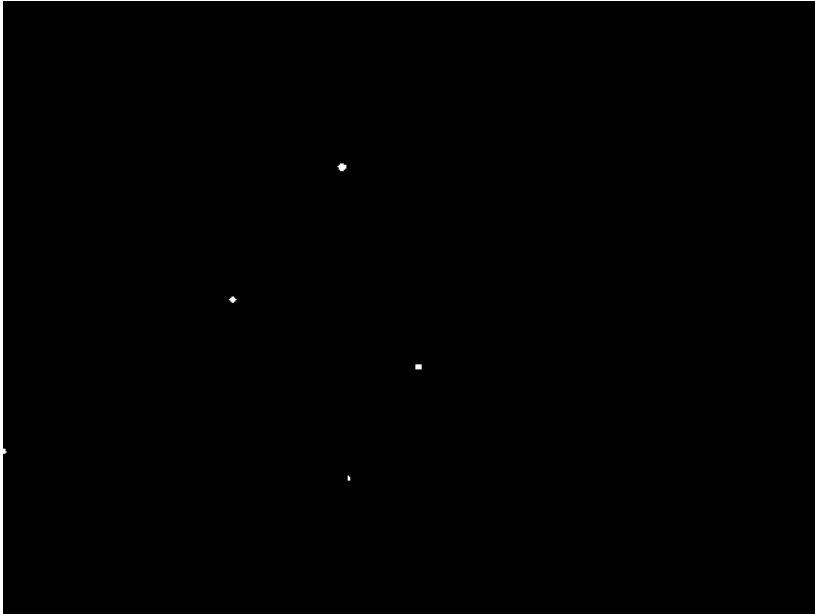
- determine the refractive index of extracellular vesicles <500 nm in suspension

# Methods - setup

- Commercial instrument
  - Nanosight NS-500



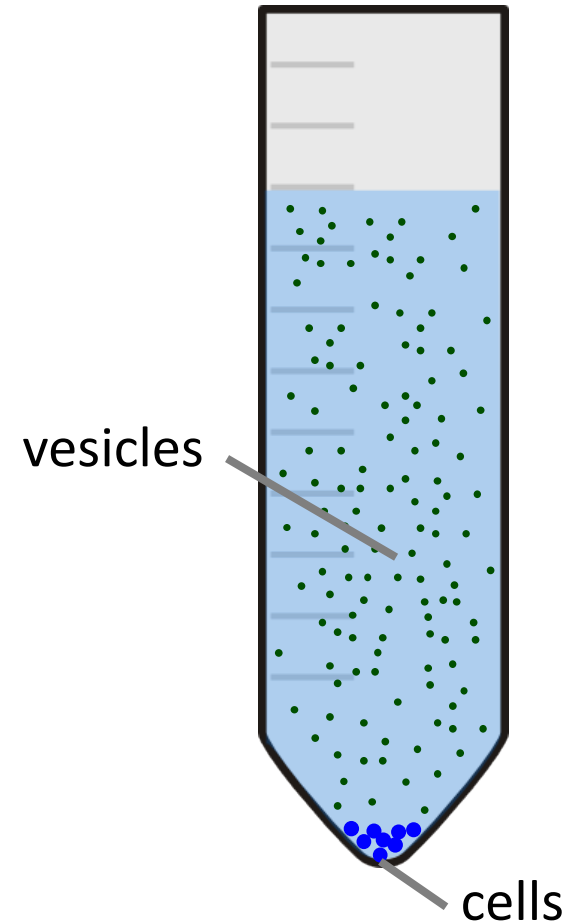
# Methods – NTA



- obtain **particle diameter**  $d$  by tracking the Brownian motion of single particles (Stokes-Einstein equation)
- measure **scattering power**  $P$
- derive particle **refractive index**  $n(P,d)$  from Mie theory

# Methods - samples

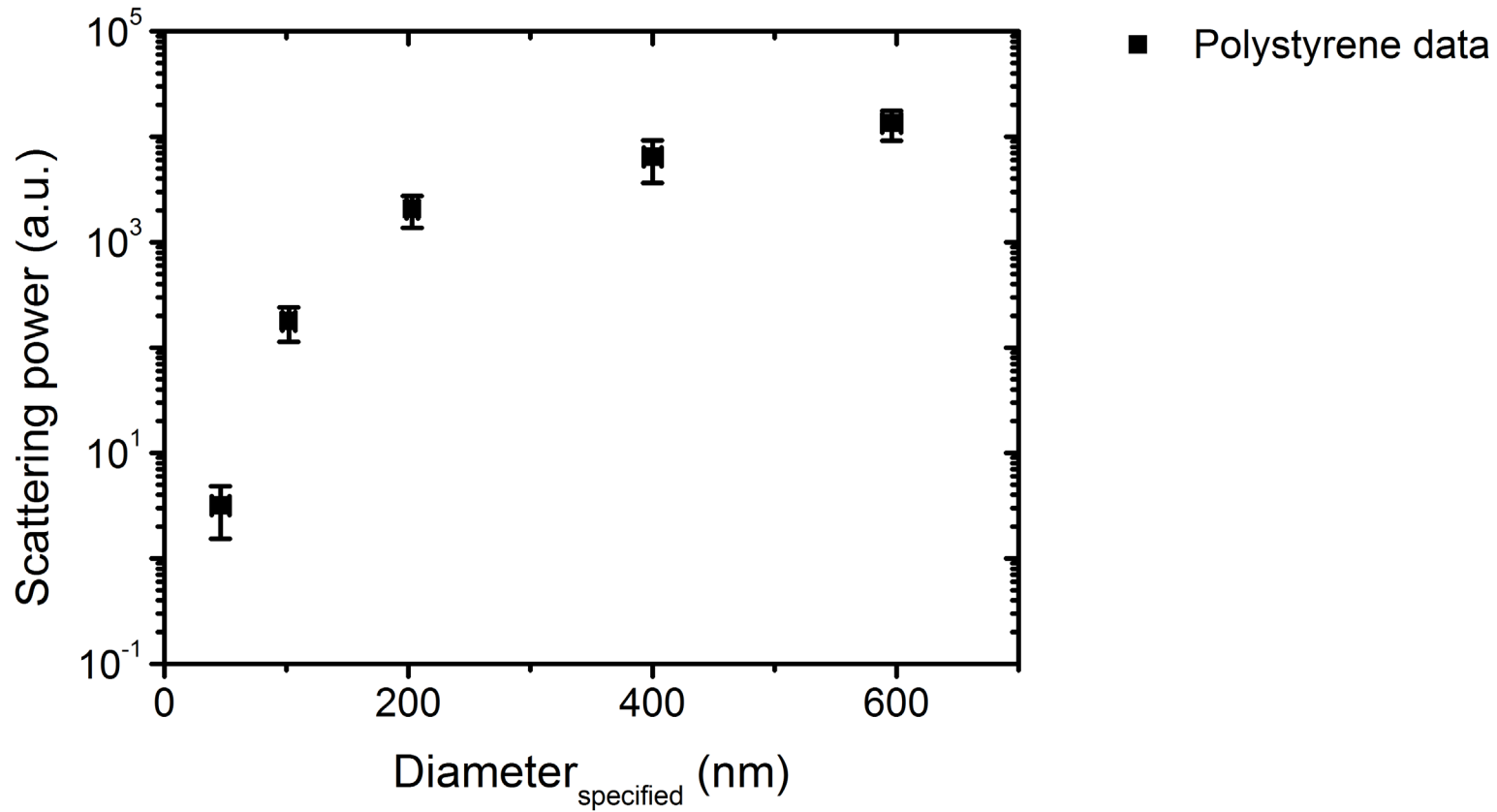
- Polystyrene beads ( $n=1.63$ )
  - Thermo Fisher Scientific, USA
- Silica beads ( $n=1.45$ )
  - Kisker Biotech, Germany
- Human urinary vesicles
  - differential centrifugation
  - protocol from [metves.eu](http://metves.eu)



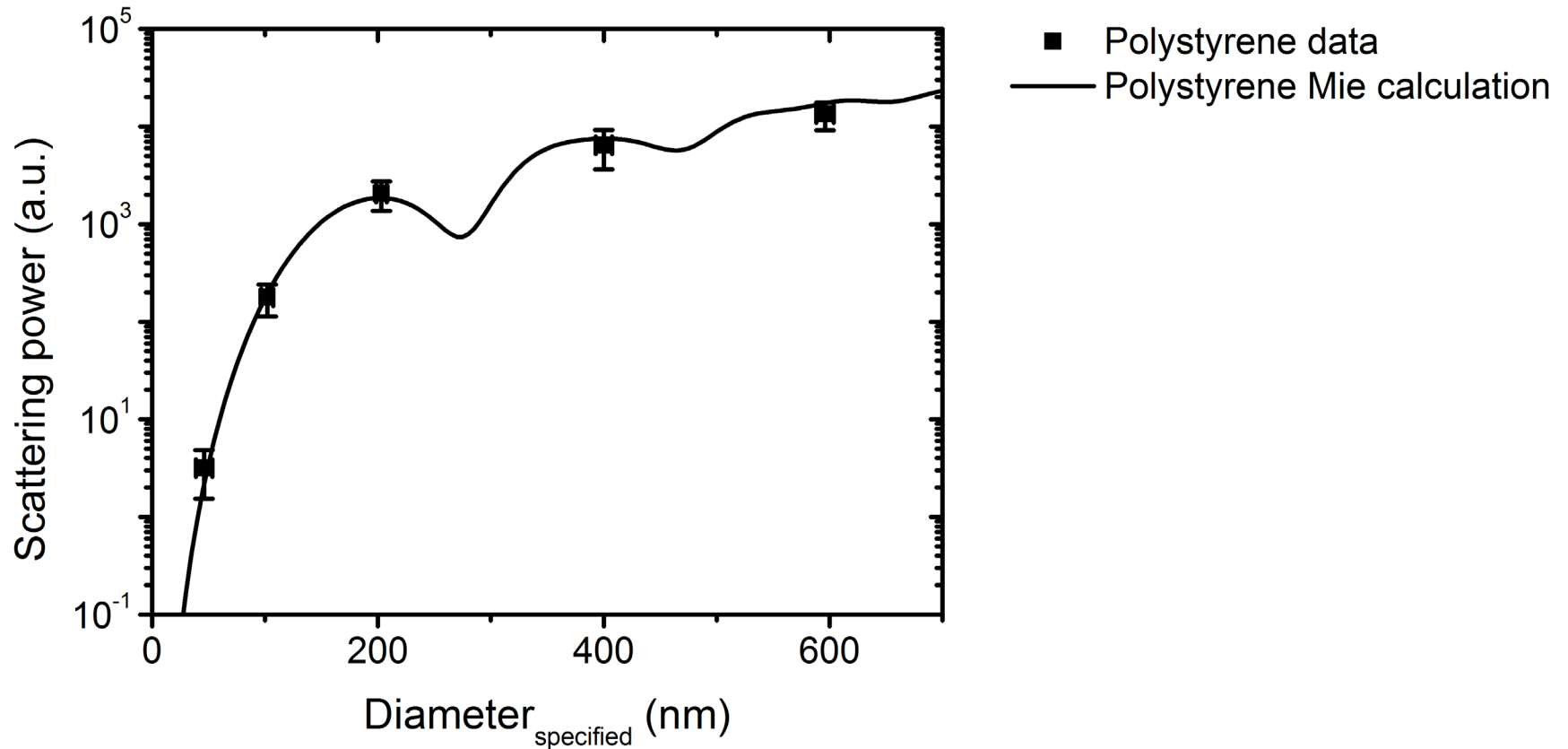
# Methods - approach

- calibration
  - measure light scattering of beads
  - describe measurements by Mie theory
- validation
  - measure light scattering and diameter of beads mixture
- application
  - determine the refractive index of vesicles

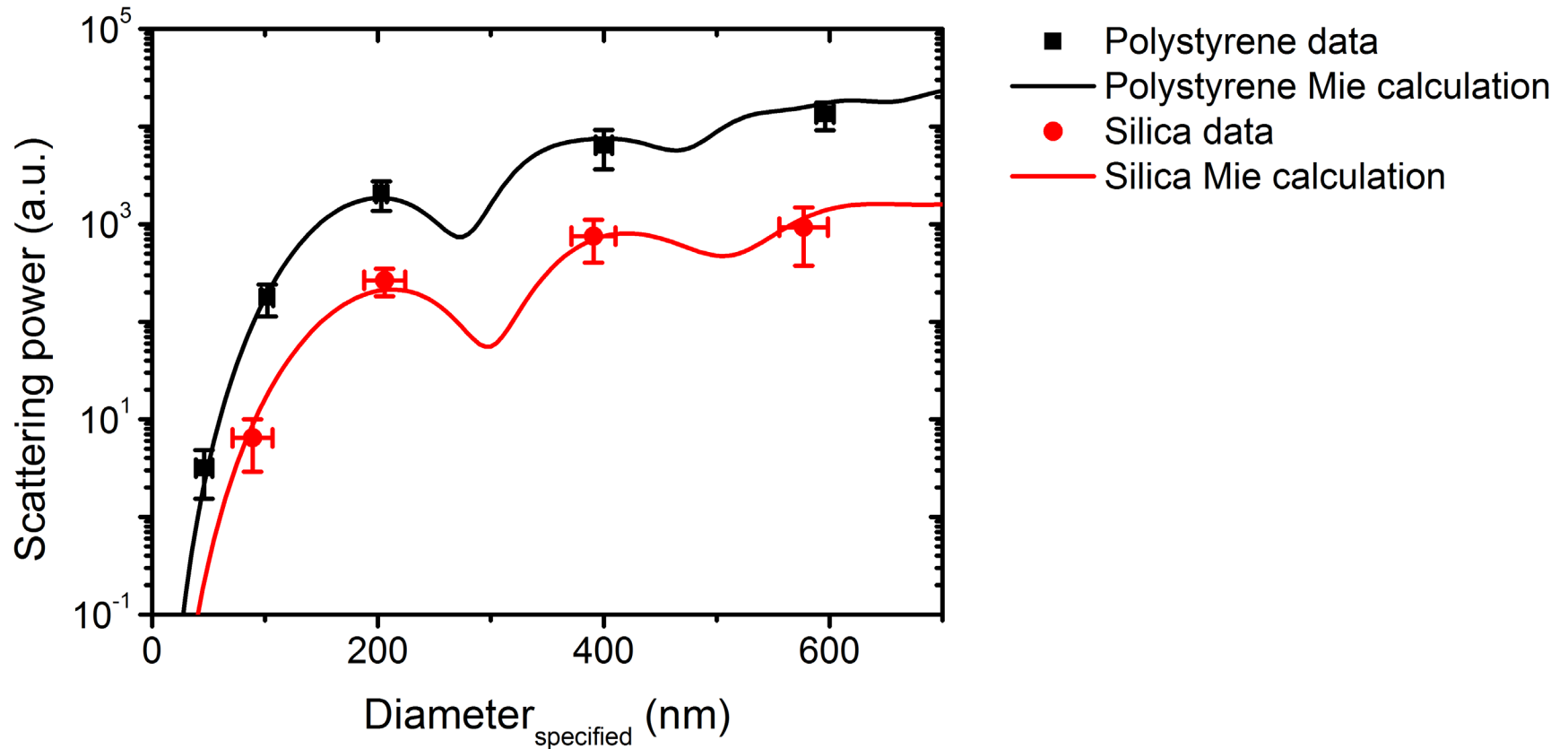
# Results - scattering power versus diameter of polystyrene beads



# Results - scattering power versus diameter of polystyrene beads described by Mie theory



# Results - scattering power versus diameter of polystyrene and silica beads





# Methods - approach

## ✓ calibration

- ✓ measure light scattering of beads
- ✓ describe measurements by Mie theory

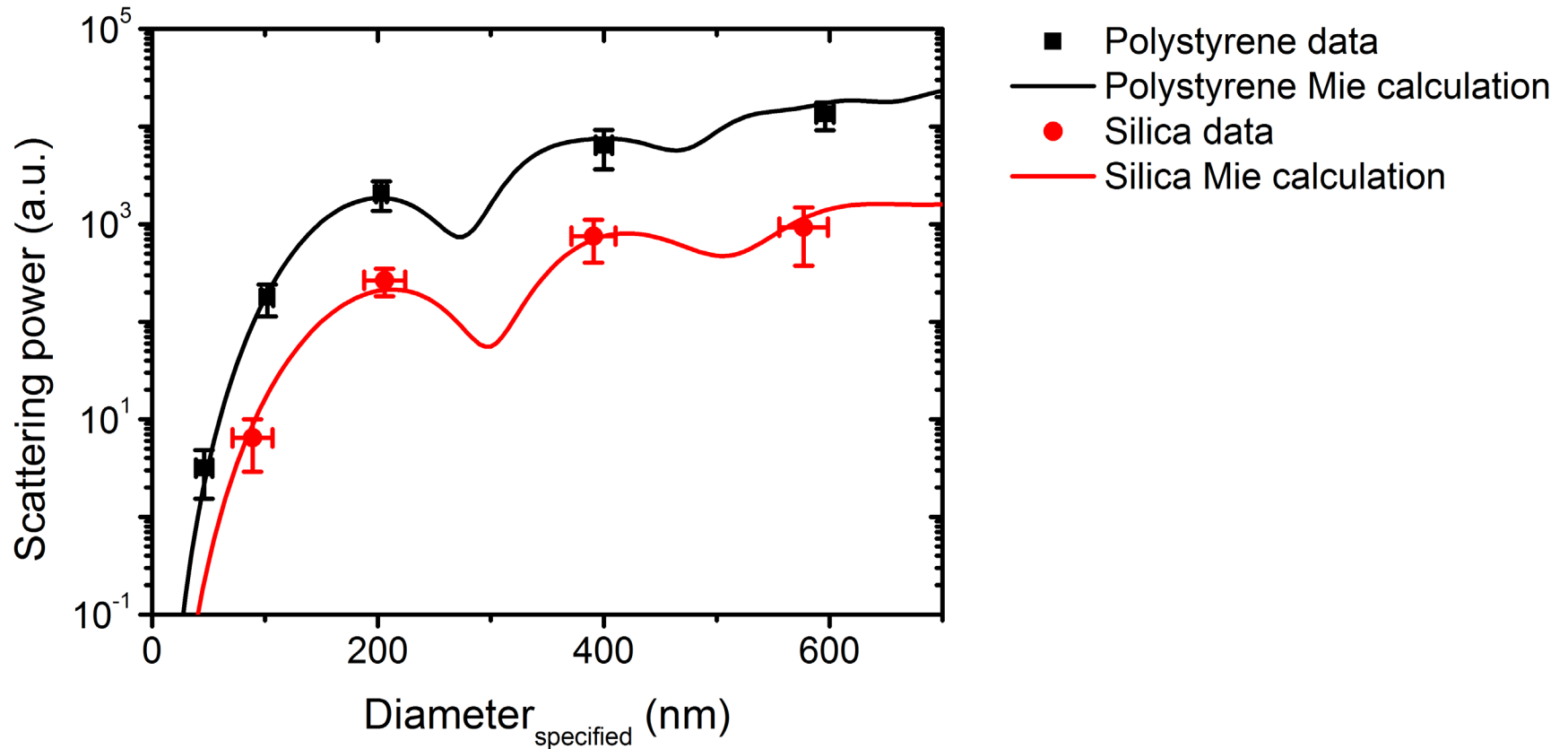
## ● validation

- measure light scattering and diameter of beads mixture

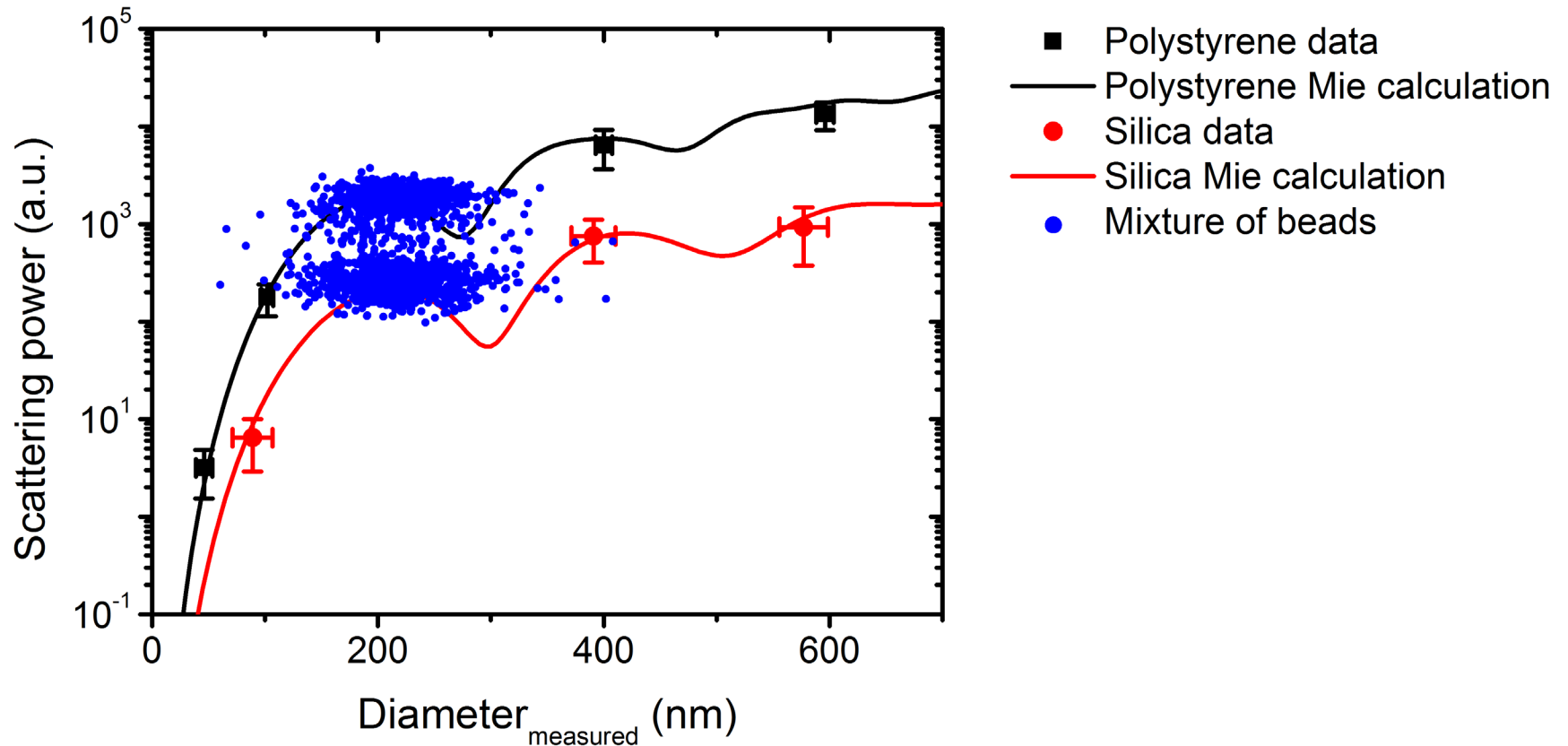
## ● application

- determine the refractive index of vesicles

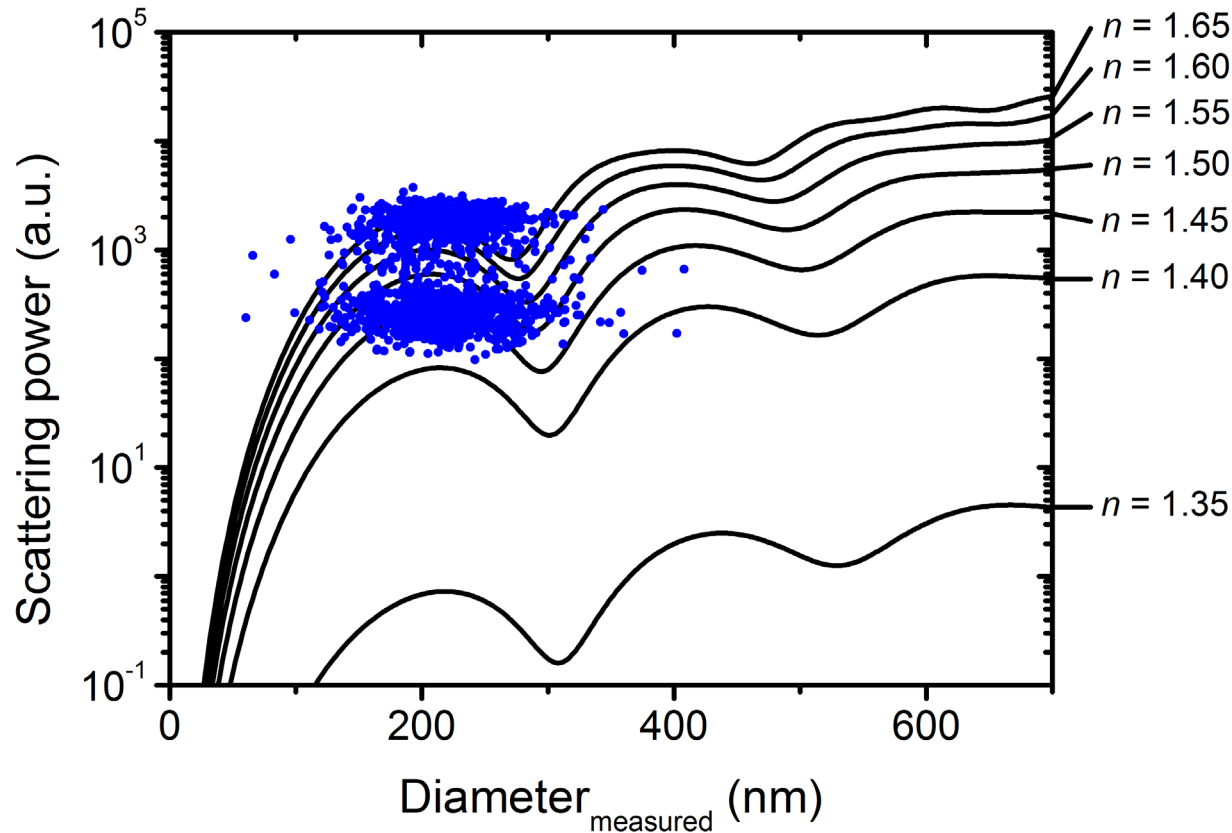
# Results - scattering power versus diameter of polystyrene and silica beads



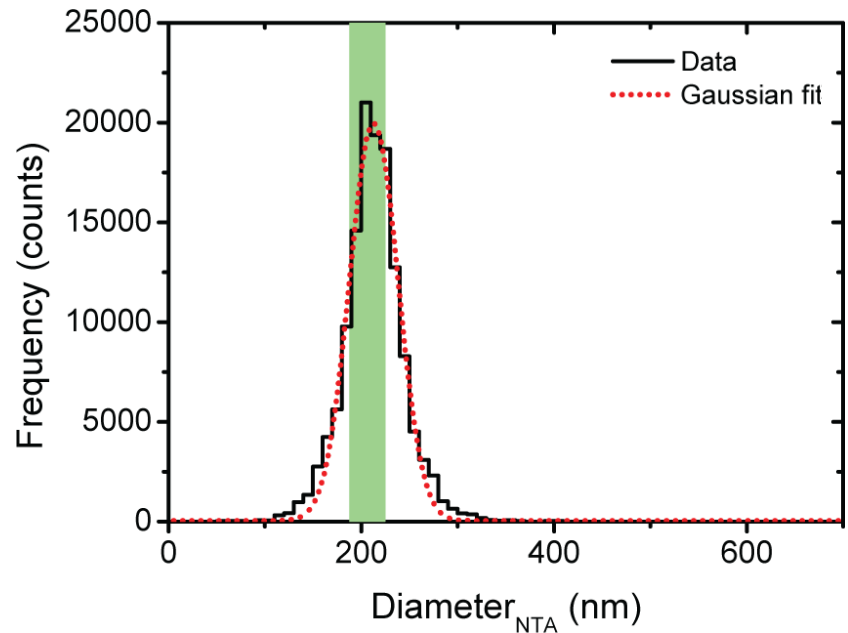
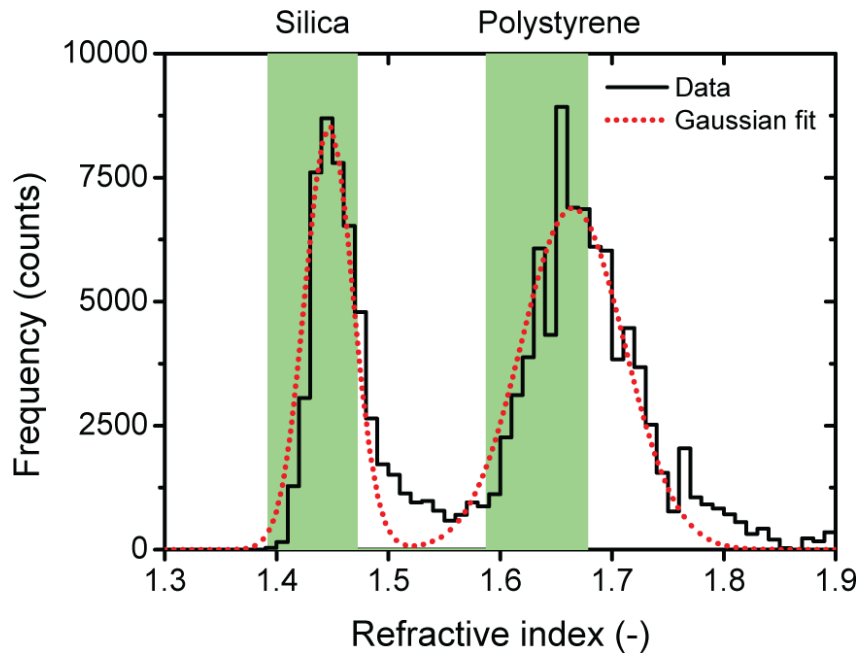
# Results - scattering power versus diameter of a mixture of polystyrene and silica beads



# Results - scattering power versus diameter of a mixture of polystyrene and silica beads



# Results - refractive index and size distribution of a mixture of polystyrene and silica beads



# Methods - approach

## ✓ calibration

- ✓ measure light scattering of beads
- ✓ describe measurements by Mie theory

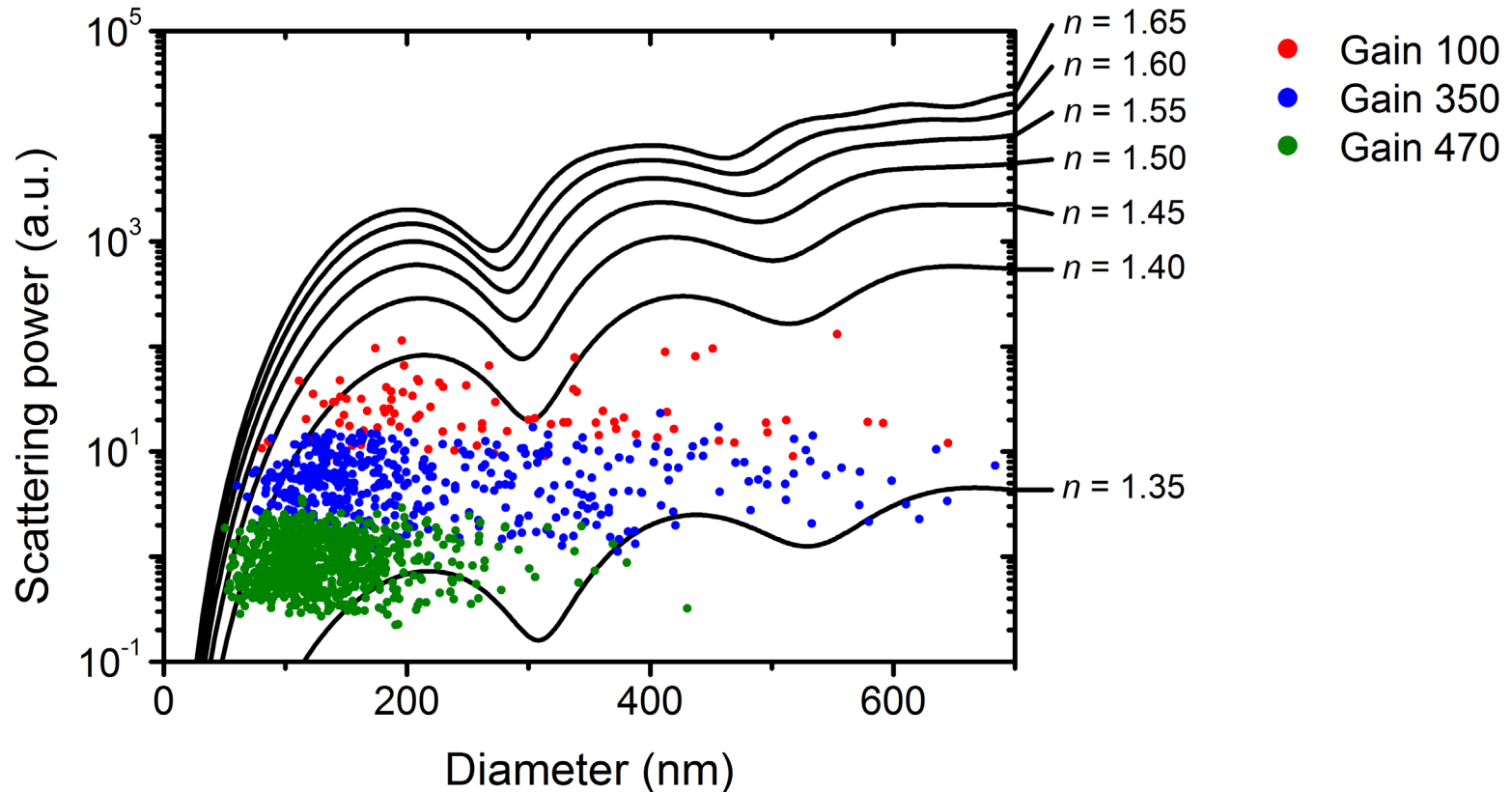
## ✓ validation

- ✓ measure light scattering and diameter of beads mixture

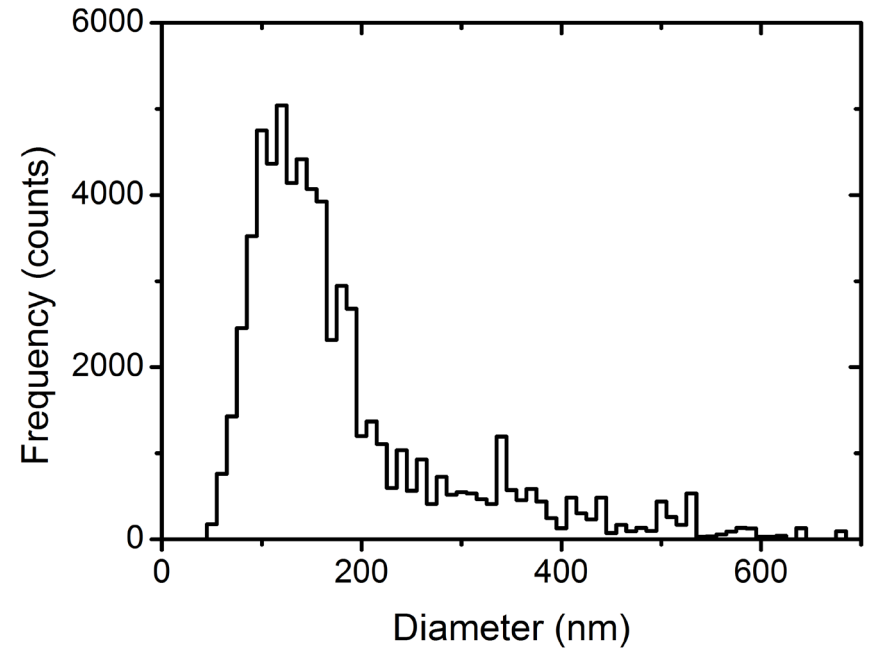
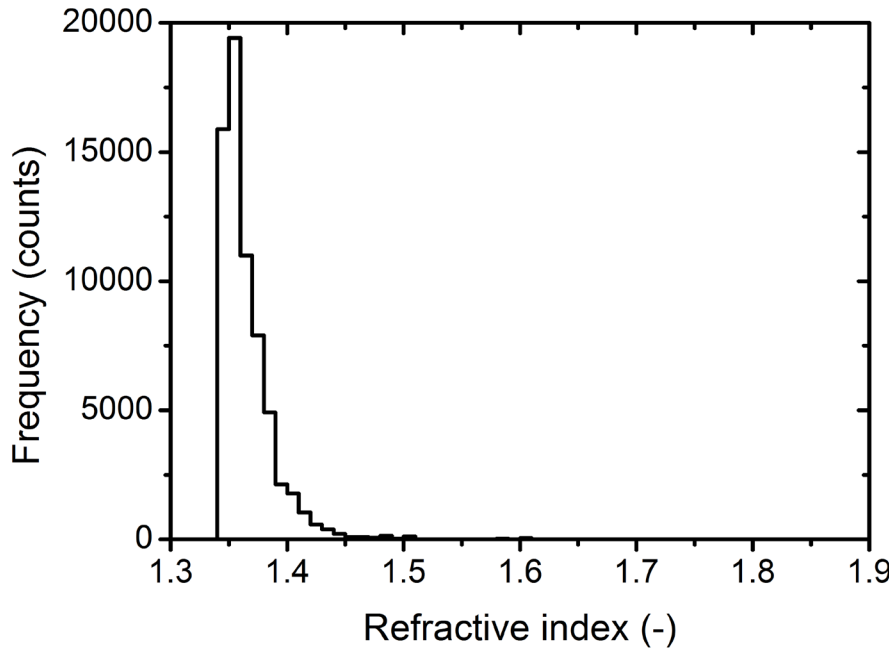
## ● application

- determine the refractive index of vesicles

# Results - scattering power versus diameter of urinary vesicles

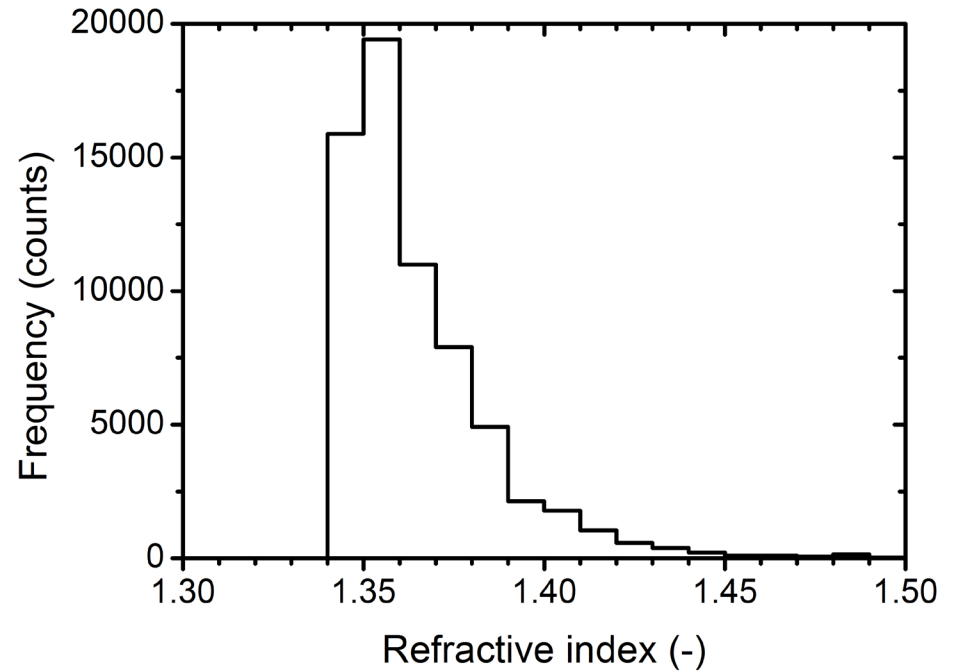


# Results - size and refractive index distribution of urinary vesicles





# Conclusions



- NTA can be used to determine the refractive index of single vesicles
- mean refractive index of urinary vesicles is 1.37

# Timeline vesicle detection with optical methods

Flow cytometry standardization <i>Lacroix et al.</i>	Flow cytometry calibration <i>Chandler et al.</i>	Scanning flow cytometry <i>Konokhova et al.</i>	Flow cytometry artifact <i>Nolan et al.</i>	Flow cytometry standardization by vesicle size approximation <i>Coumans et al.</i>
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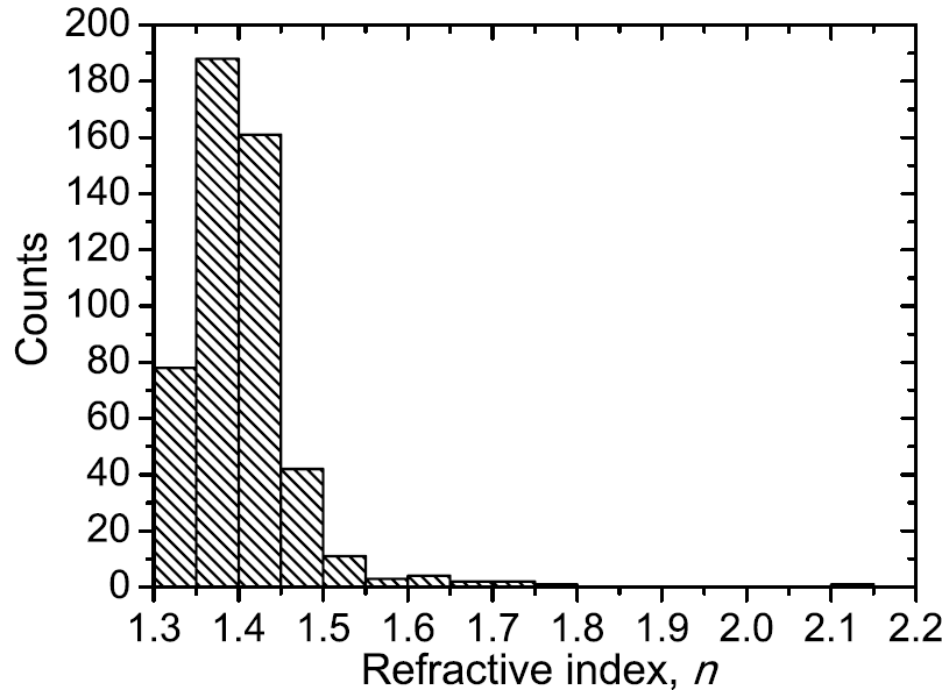
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# Refractive index of blood microparticles



- scanning flow cytometry
- microparticles >500 nm

# Timeline vesicle detection with optical methods

Flow cytometry  
standardization  
*Lacroix et al.*

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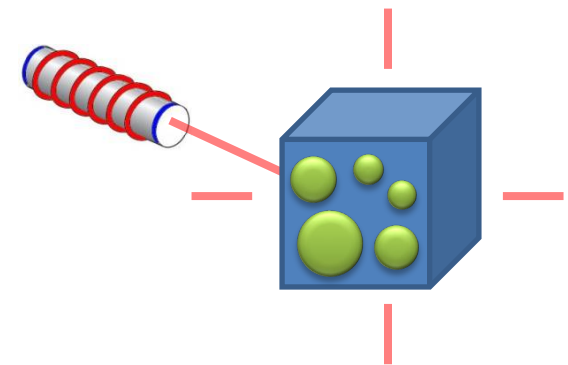
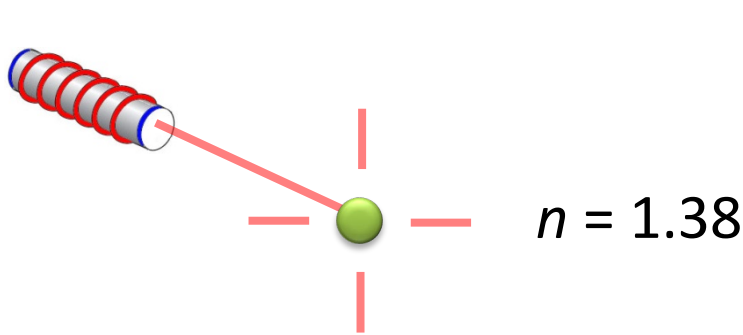
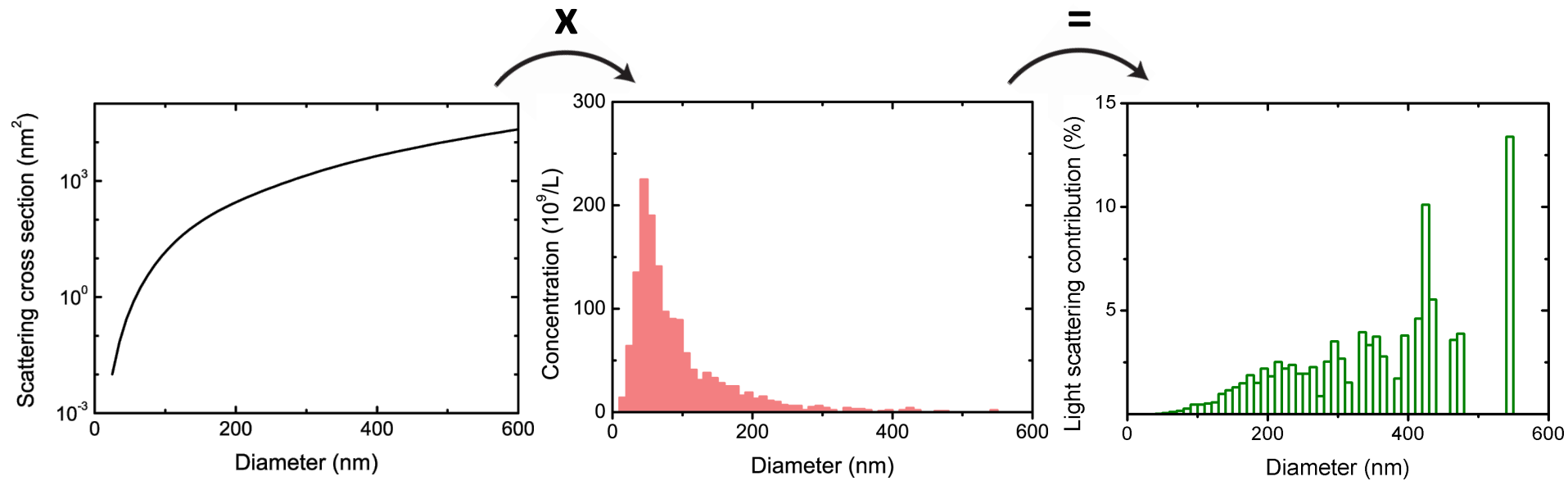
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*al.*

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# Large vesicles dominate light scattering signal



# Timeline vesicle detection with optical methods

Flow cytometry  
standardization  
*Lacroix et al.*



Flow cytometry  
*Bochkova et al.*  
Nanoman on  
vesicles  
*Scheff et al.*

Flow cytometry  
artifact  
*Nolan et al.*

Flow cytometry  
standardization  
by vesicle size  
approximation  
*Coumans et al.*

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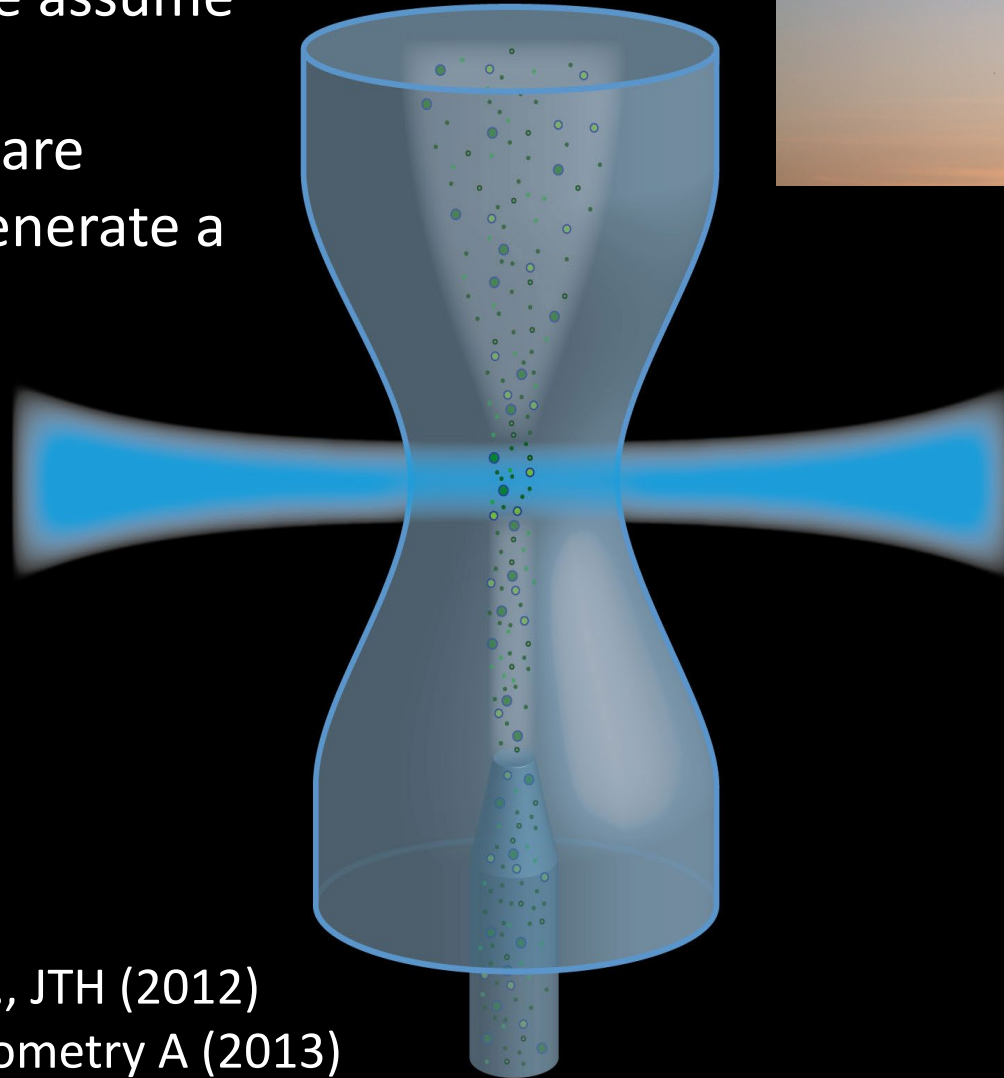
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# Swarm detection

for vesicles we assume  
 $n = 1.38$   
>150 vesicles are  
required to generate a  
signal event



van der Pol et al., JTH (2012)  
Nolan et al., Cytometry A (2013)

# Timeline vesicle detection with optical methods

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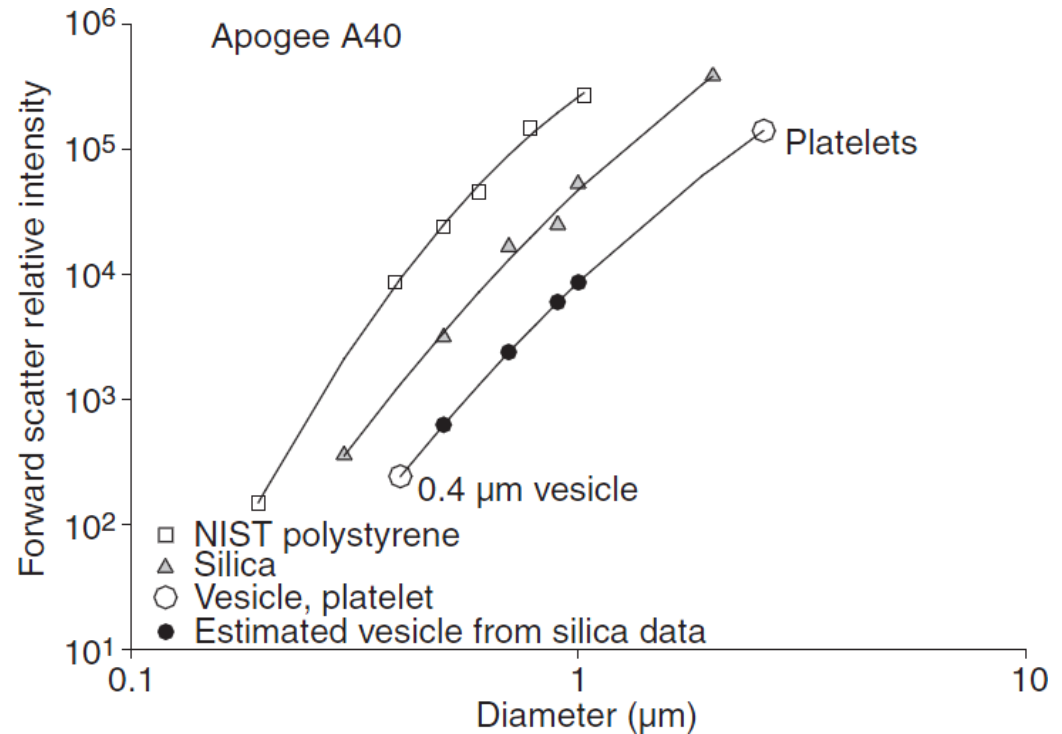
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# Enhanced insight in flow cytometry data

“The use of plastic beads for size calibration in flow cytometry remains an imperfect model. Many factors other than size influence scatter, among which are the refractive index...”



Lacroix et al.,  
Semin. Thromb. Hemost. (2010)

Chandler et al., JTH (2011)

# Timeline vesicle detection with optical methods

Flow cytometry  
standardization  
*Lacroix et al.*

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*Konokhova et al.*

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*Tatischeff et al.*

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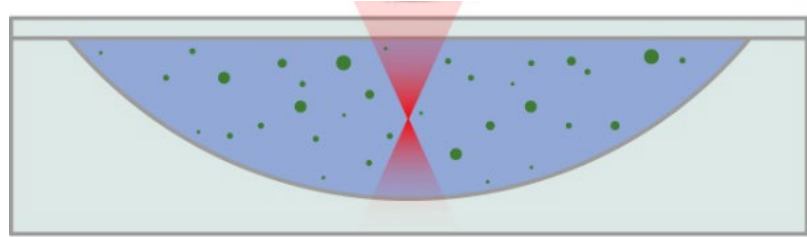
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# Refractive index affects smallest detectable size of vesicles

- Nanoparticle analysis range: typically 10 nm - 2000 nm, dependent on particle material



“Cellular vesicles have a low refractive index, and the smallest detectable size using nanoparticle tracking analysis is in the order of 50 nm....”

Dragovic et al.,  
Nanomedicine (2011)

“It is difficult to estimate whether one big vesicle or a cluster of several smaller vesicles have been optically trapped...”

Tatischeff et al.,  
J. Extracellular Vesicles (2012)

# Outlook vesicle detection with optical methods

Flow cytometry  
standardization

*Lacroix et al.*

Flow cytometry  
calibration

*Chan*

Nanovesicle  
tracking

*Drag*

Scanning flow

Flow cytometry

Flow cytometry  
standardization

by vesicle size  
approximation  
*Coumans et al.*

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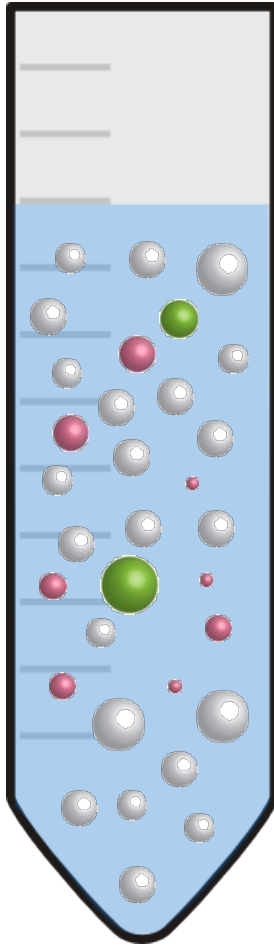
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# Refractive index enables flow cytometry standardization



# Distinguish vesicles from other particles



- lipoproteins ( $n = 1.45-1.60$ )
- protein aggregates ( $n = 1.53-1.60$ )
- vesicles ( $n < 1.45$ )



# Refractive index reveals vesicle composition

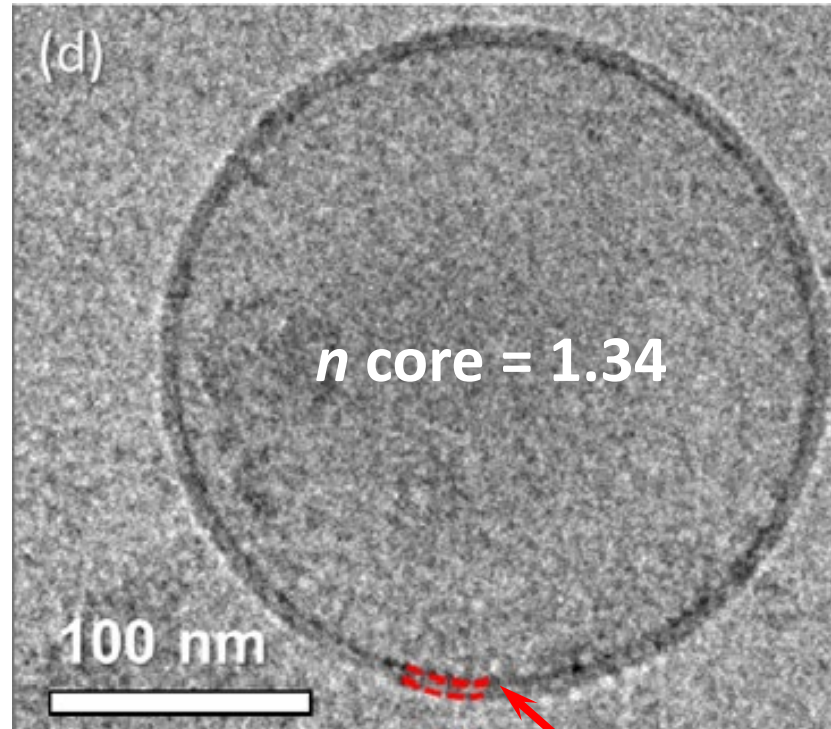
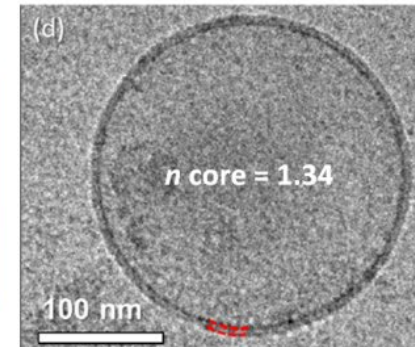
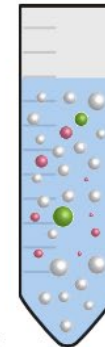
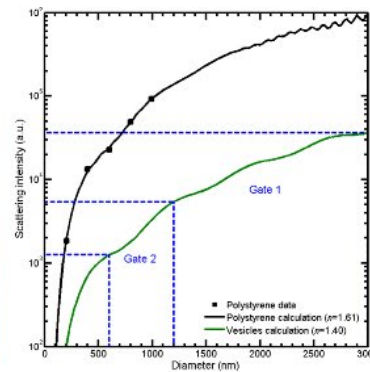
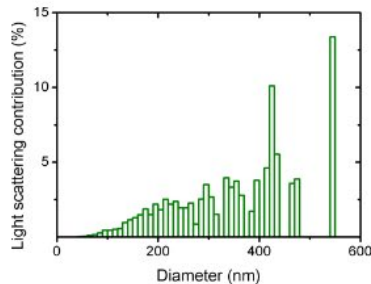


image courtesy of Issman et al., PLoS ONE (2013)

\* van Manen et al., Biophys. J. (2007)

# Summary: implications of the vesicle refractive index

- insight
  - large vesicles dominate scattering
  - swarm detection
- relate scatter to size
  - detection limit
  - flow cytometry standardization
- vesicle identification
- vesicle composition





# Acknowledgements

- Academic Medical Center
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  - Chris Gardiner
- University of Birmingham
  - Paul Harrison
- NanoSight Ltd.
  - Andrew Malloy
  - Patrick Hole
  - Jonathan Smith

More on vesicle detection:  
[edwinvanderpol.com](http://edwinvanderpol.com)

