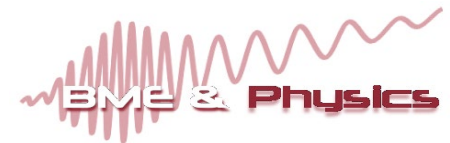


# Trapping of single extracellular vesicles in the evanescent field of an optical cavity

Edwin van der Pol<sup>1,2</sup>

Frank Coumans<sup>1,2</sup>, J. Wilke<sup>3</sup>, C. Earhart<sup>3</sup>, B. DiPaolo<sup>3</sup>,  
R. Hart<sup>3</sup>, B. Cordovez<sup>3</sup>, Auguste Sturk<sup>2</sup>,  
Rienk Nieuwland<sup>2</sup>, and Ton van Leeuwen<sup>1</sup>

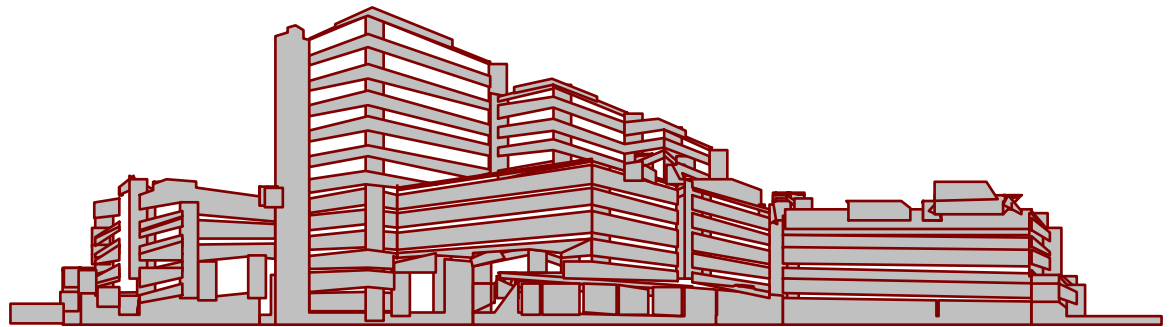
February 9<sup>th</sup>, 2015



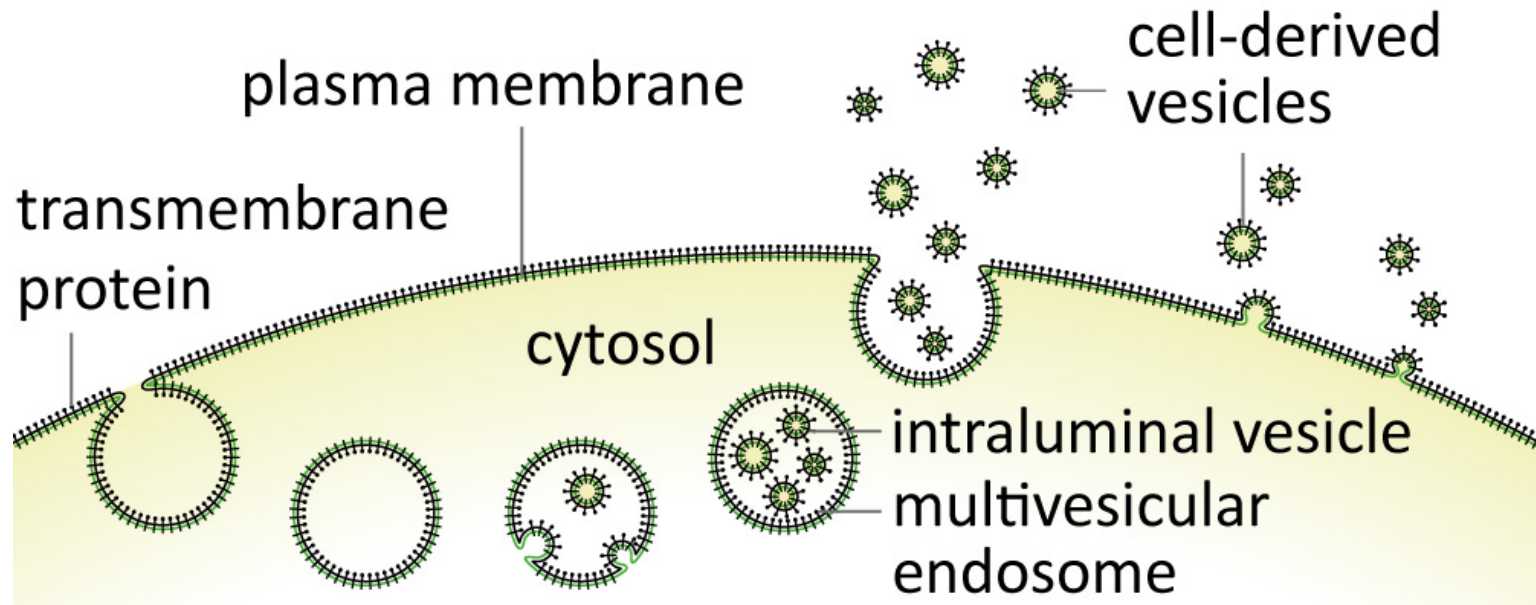
*<sup>1</sup>Biomedical Engineering and Physics; <sup>2</sup>Laboratory Experimental Clinical Chemistry, Academic Medical Center, Amsterdam, The Netherlands; <sup>3</sup>Optofluidics Inc, Philadelphia, United States of America*

# Acknowledgements

- Academic Medical Center
  - Biomedical Engineering and Physics
  - Laboratory Experimental Clinical Chemistry
- University of Twente
  - Aufried Lenferink
  - Cees Otto
- Optofluidics
  - Bernardo Cordovez

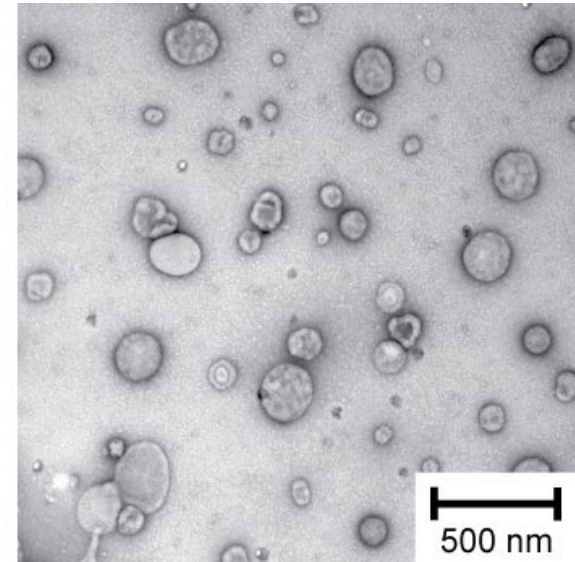
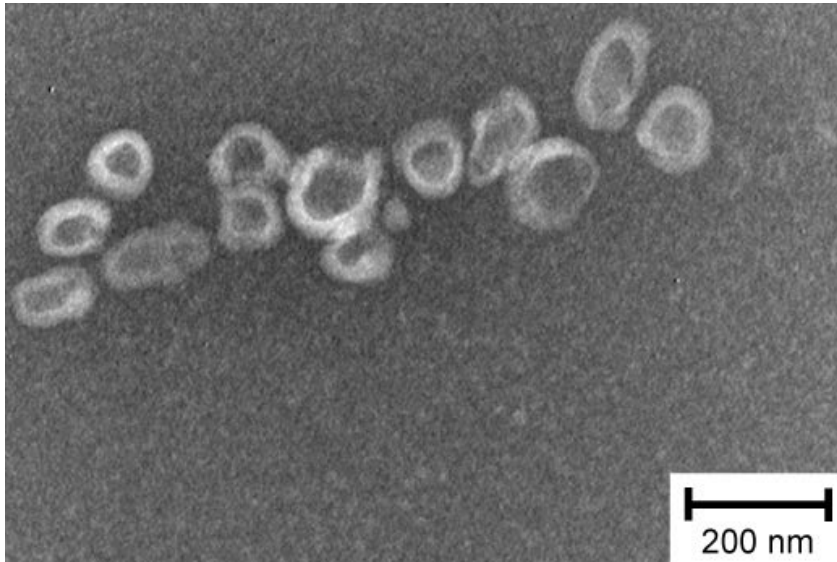


# Introduction – extracellular vesicles



- cells release vesicles (e.g. exosomes):  
biological nanoparticles with receptors, DNA, RNA
- specialized functions
- clinically relevant

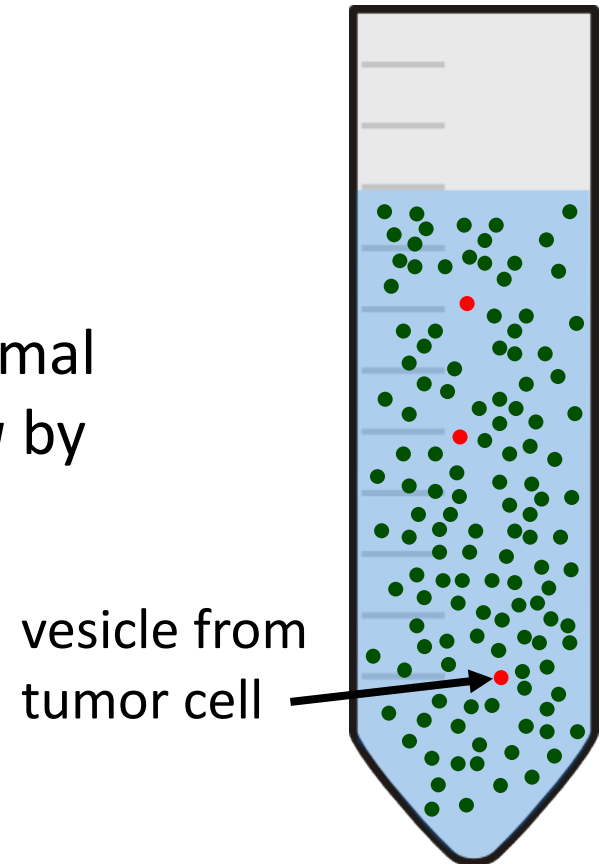
# Introduction – extracellular vesicles



- vesicle detection is cumbersome
  - small size (< 500 nm)
  - low refractive index (~1.4)
  - fluorescent antibody labeling involves practical problems

# Motive and goal

- Clinical motive
  - count tumor vesicles in blood for therapy monitoring
- Goal
  - distinguish tumor vesicles from normal vesicles in solution *without labeling* by Raman microspectroscopy



# Sample – vesicle isolation



erythrocyte vesicles

- centrifuge (3·20 min, 1560·g) \*



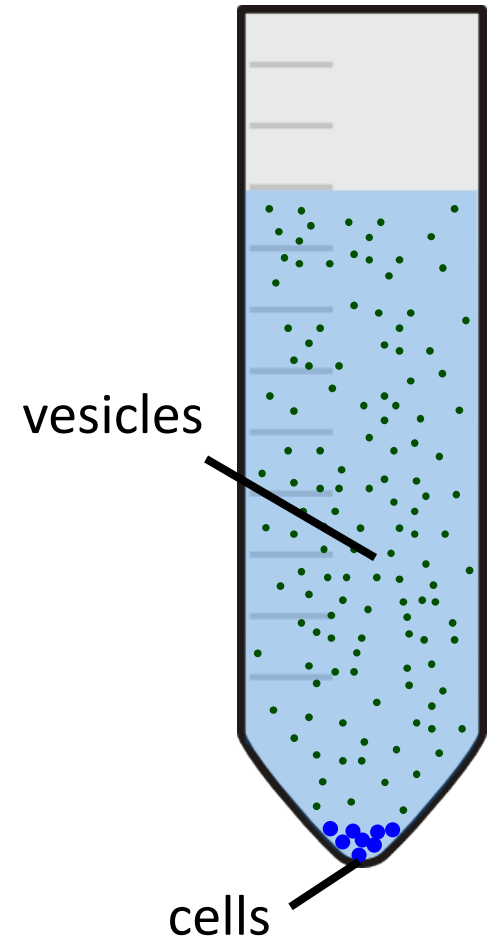
platelet vesicles

- centrifuge (3·20 min, 800·g)

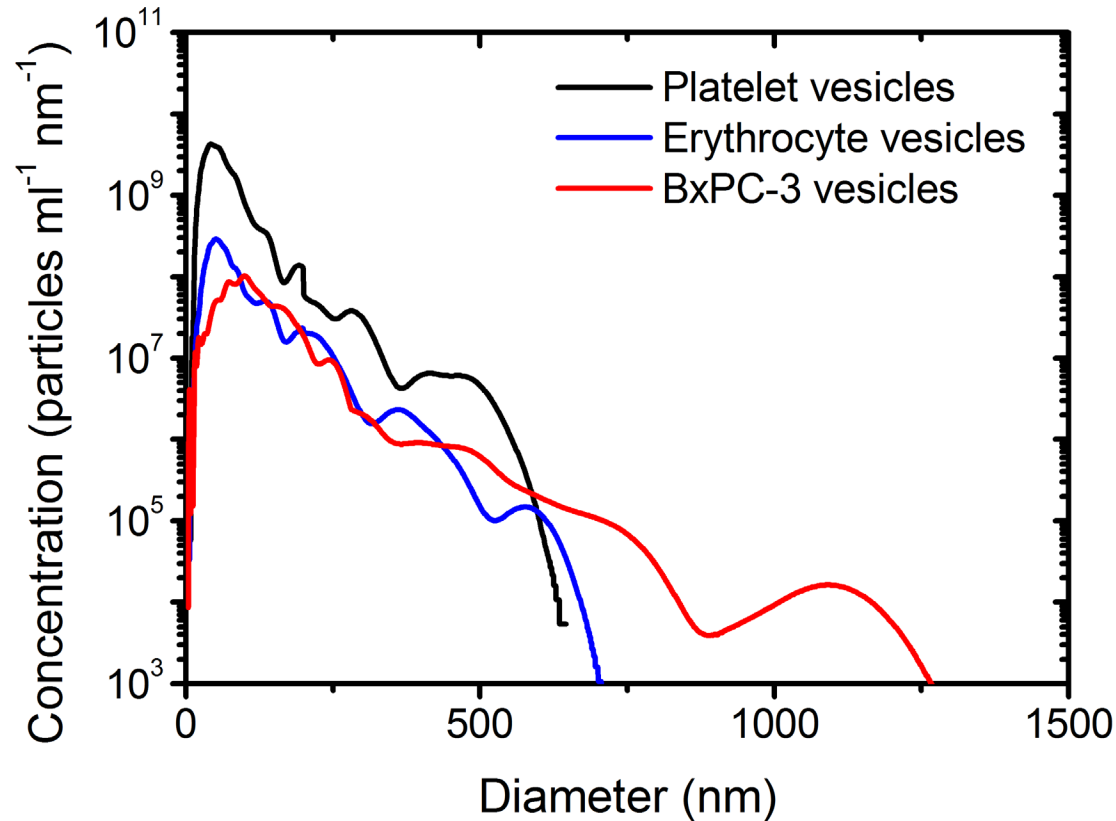


tumor vesicles from a human  
pancreatic adenocarcinoma  
(BxPC-3) cell line

- centrifuge (10 min, 180·g)

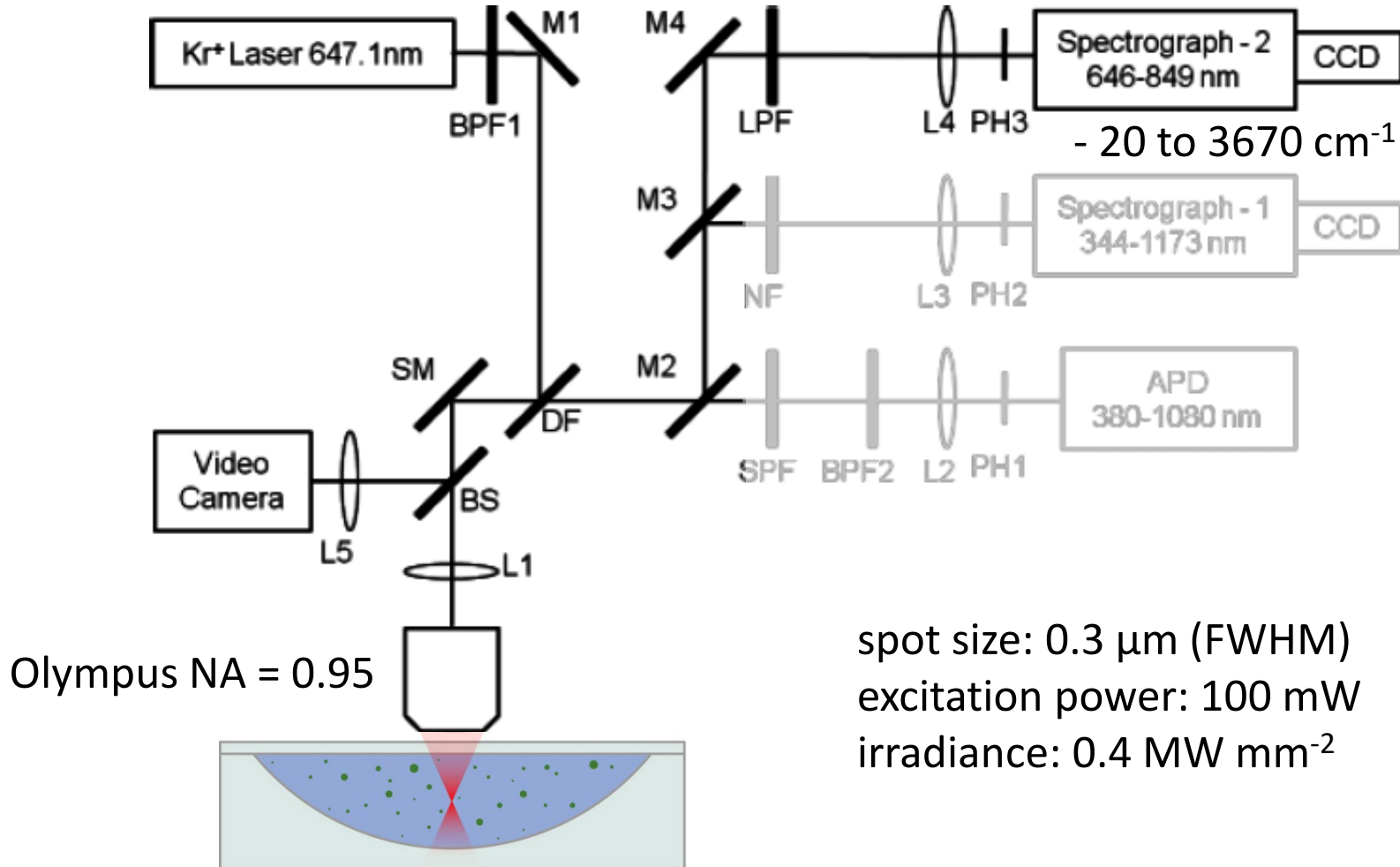


# Sample – Vesicle size distribution



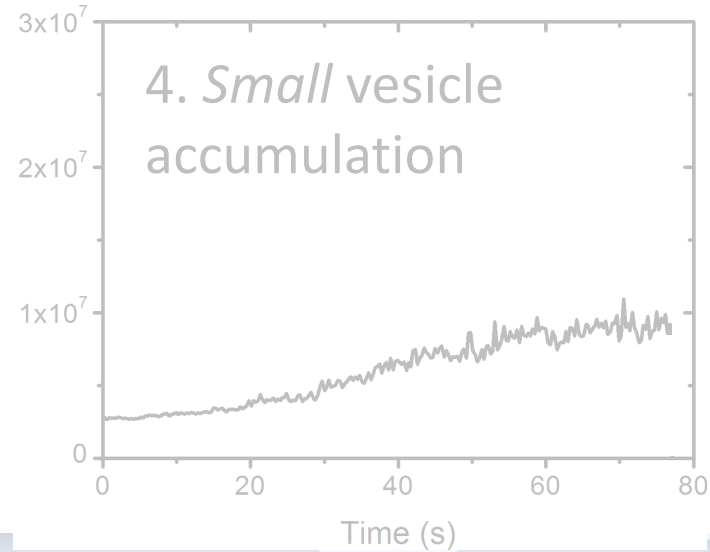
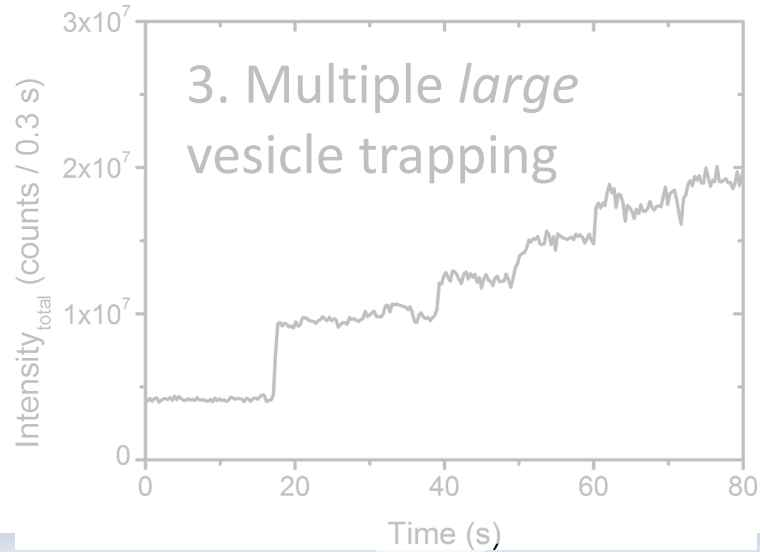
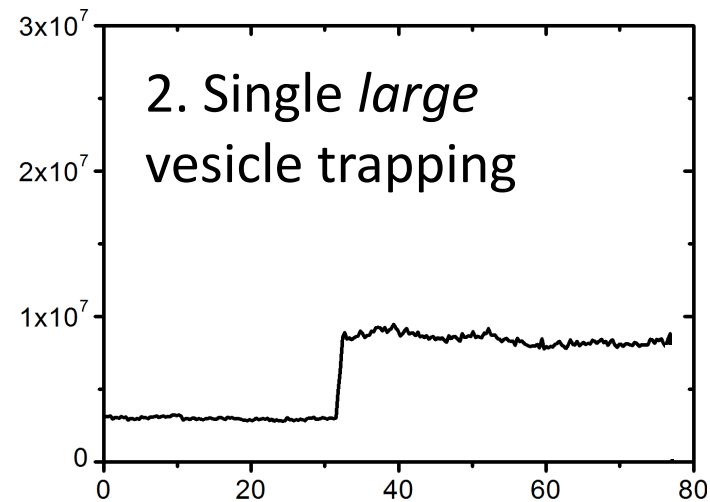
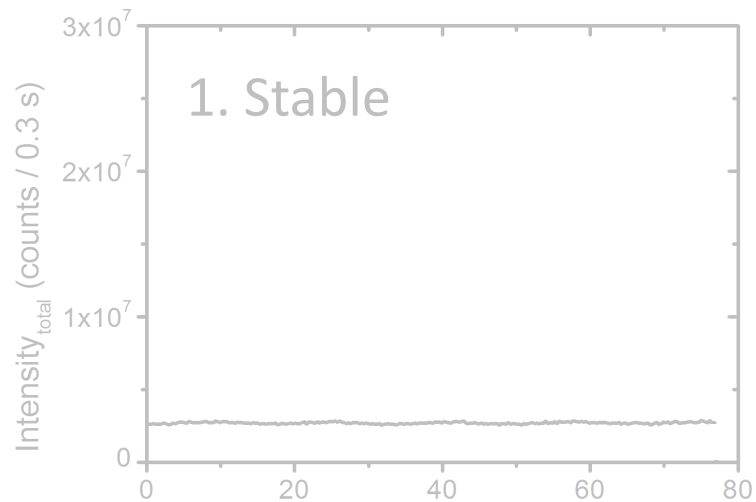
- data obtained with nanoparticle tracking analysis

# Methods – Raman microspectroscopy

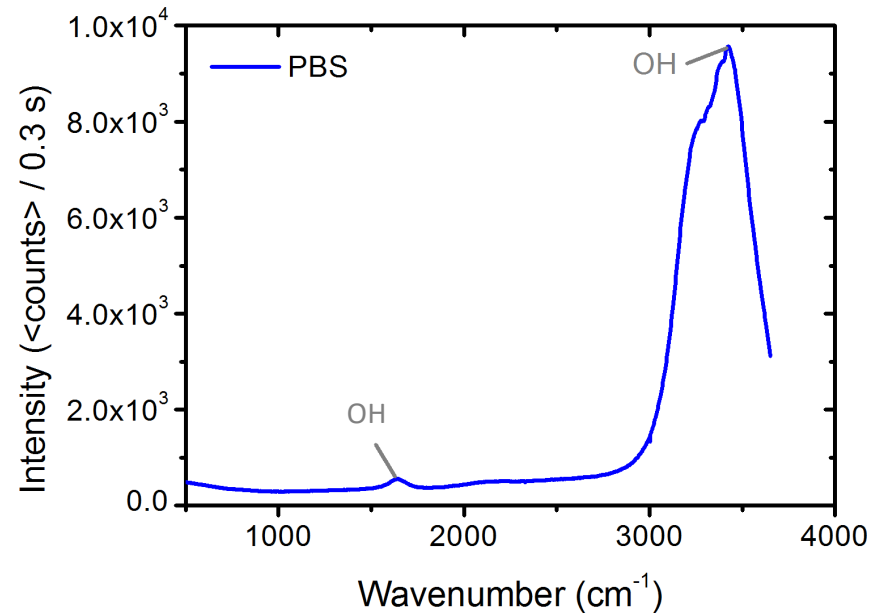
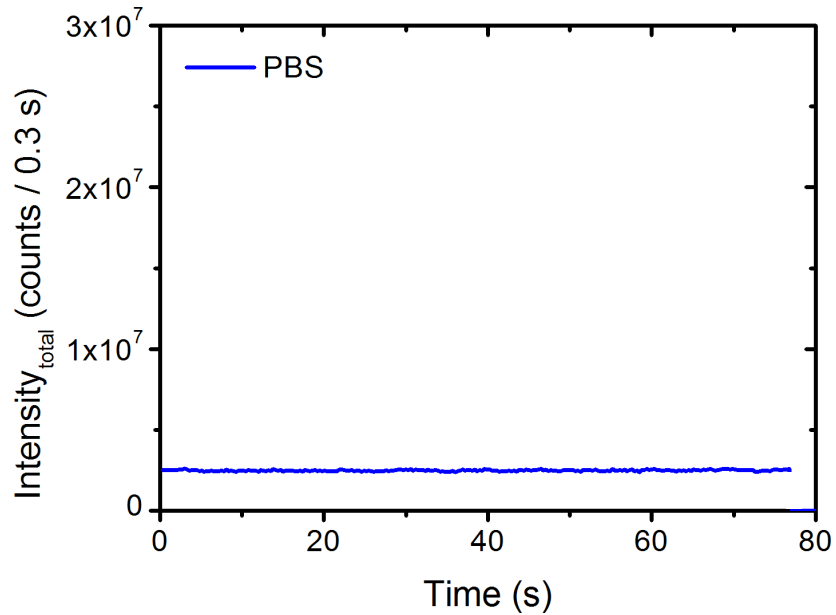




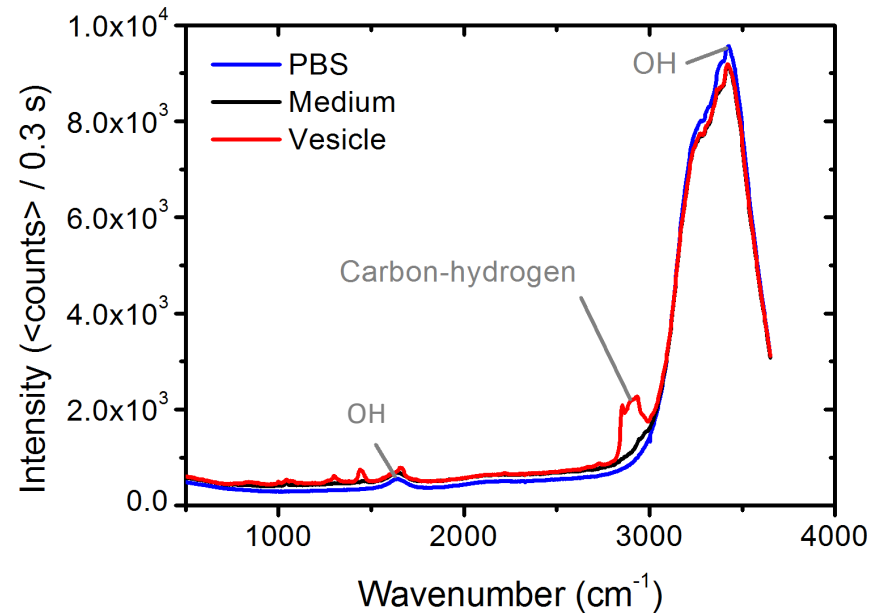
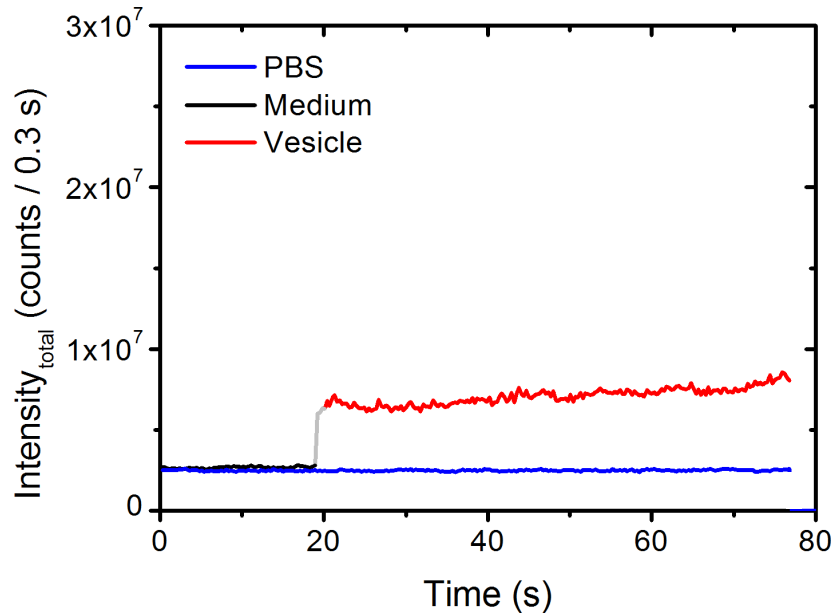
# Results – Total intensity versus time



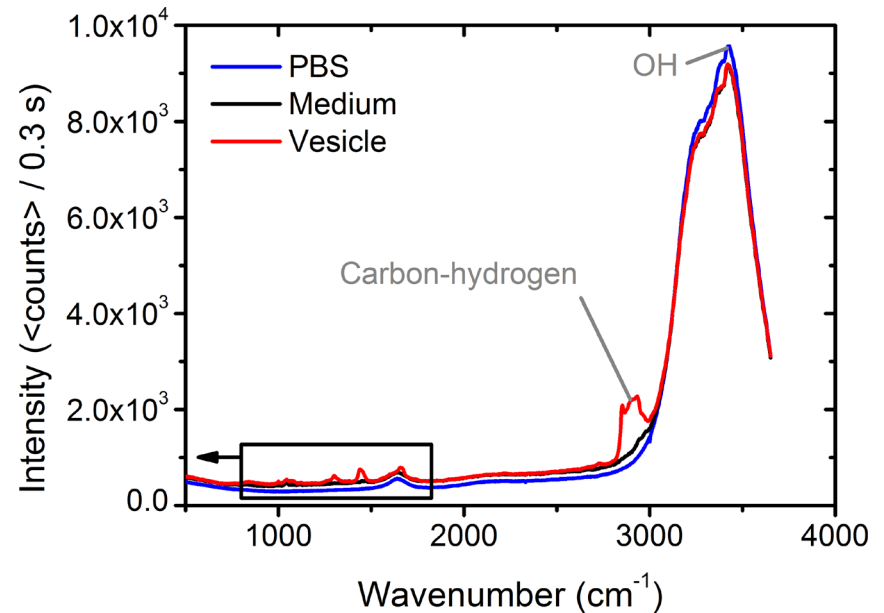
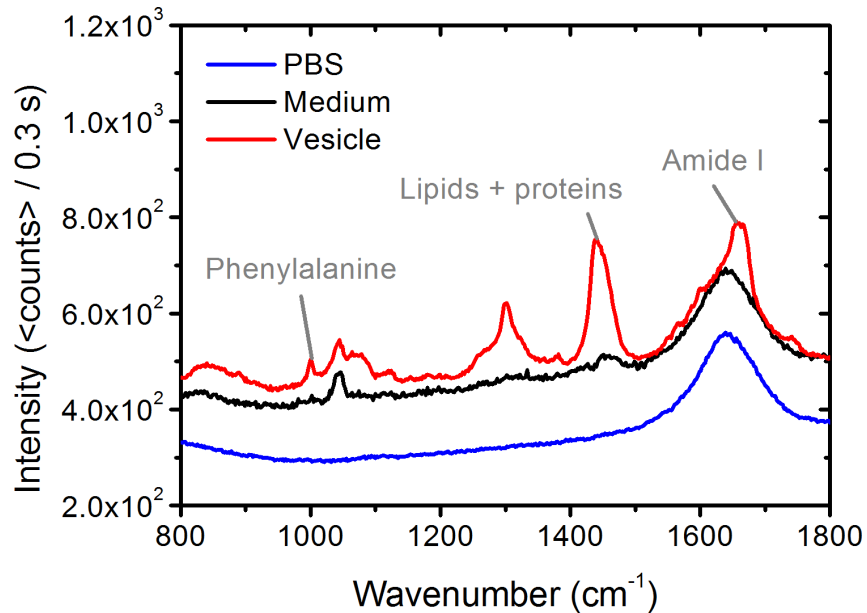
# Results - Raman spectrum of PBS



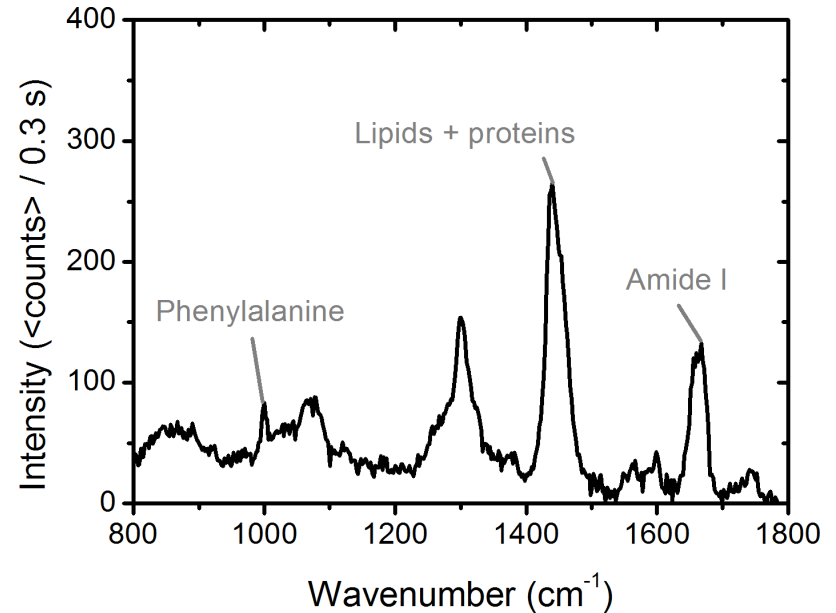
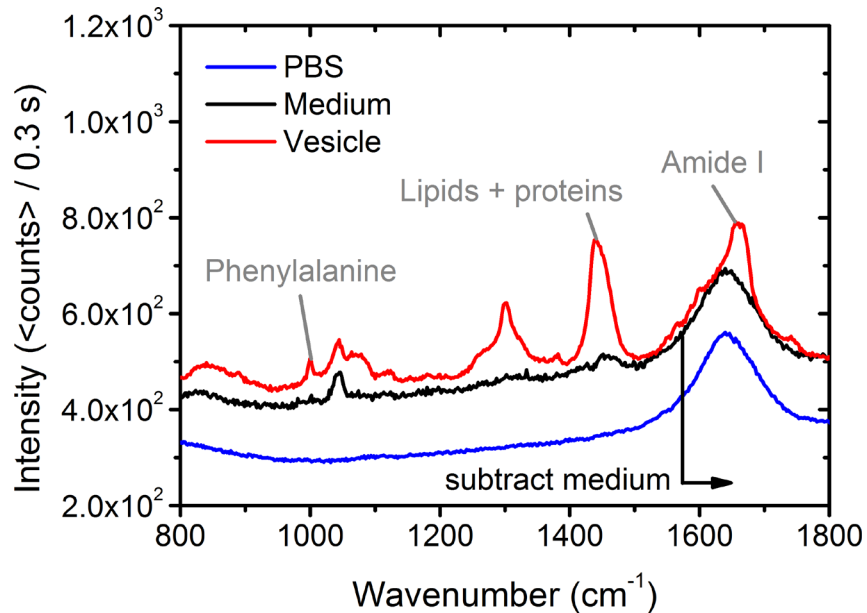
# Results - Raman spectrum of *single* tumor (BxPC-3) vesicle



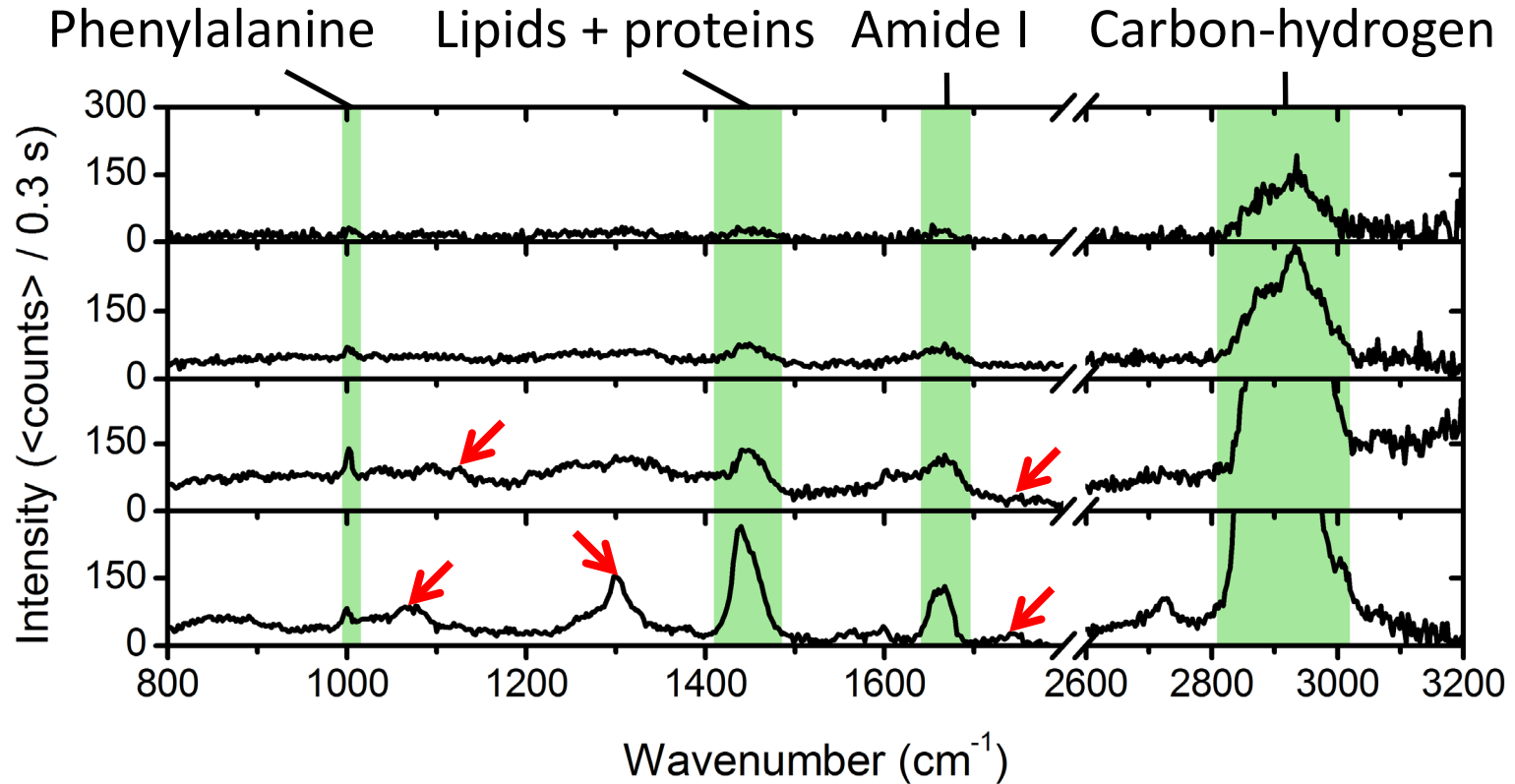
# Results - Raman spectrum of *single tumor (BxPC-3) vesicle*



# Results - Raman spectrum of *single tumor (BxPC-3) vesicle*

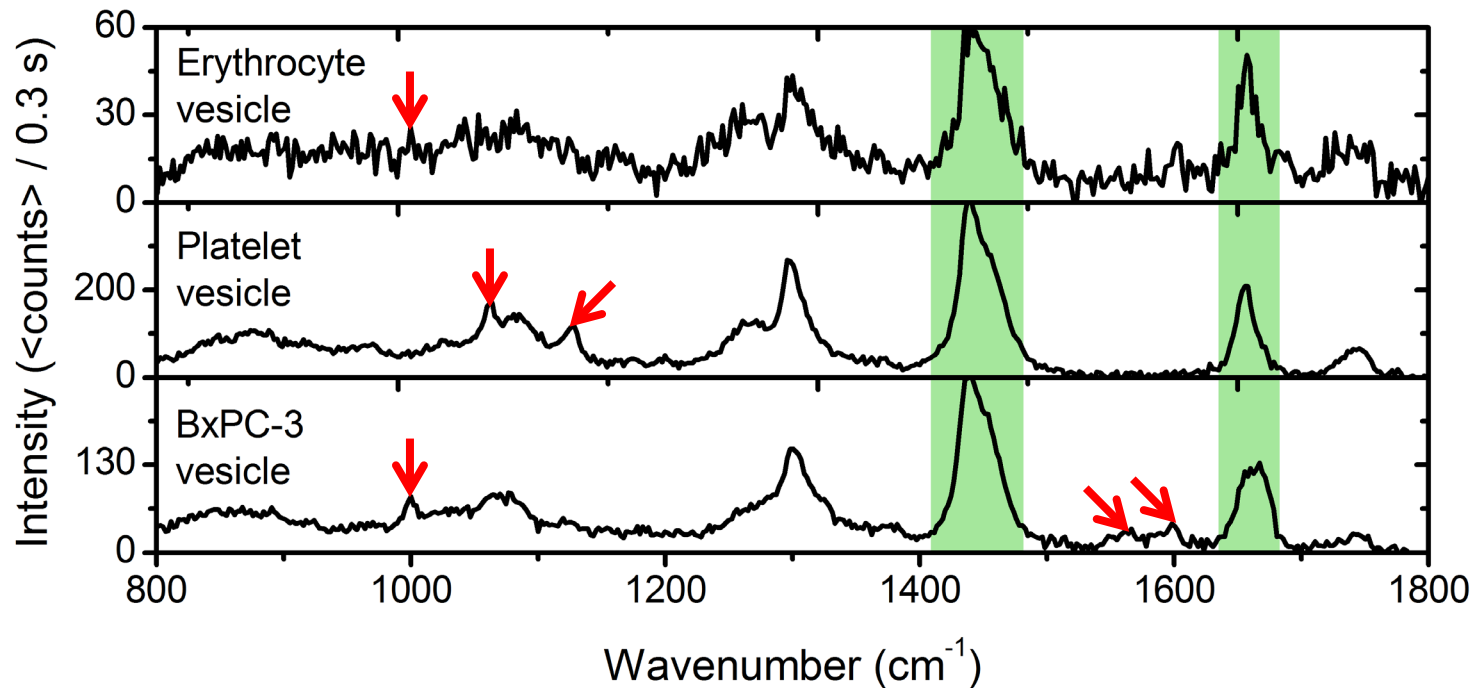


# Results – Single BxPC-3 vesicle trapping



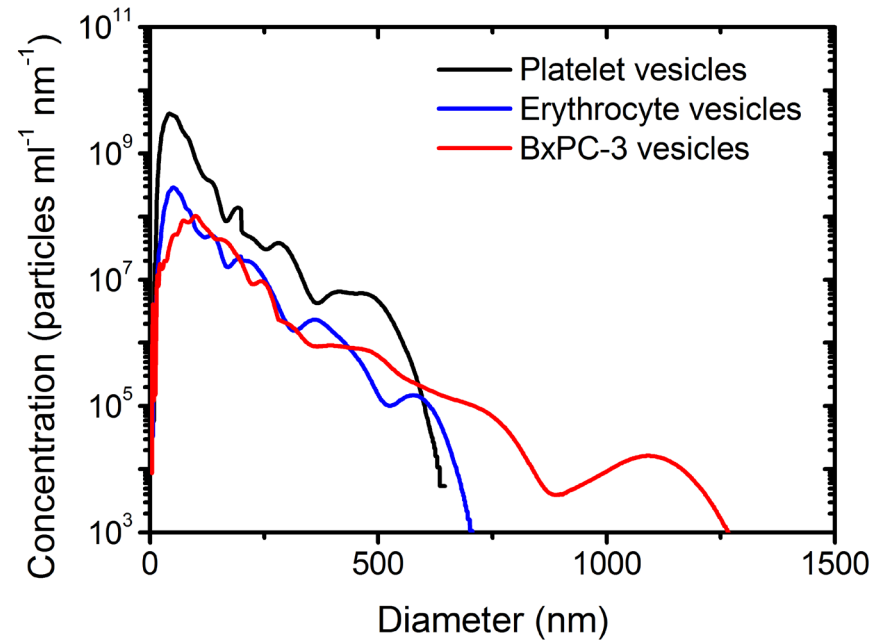
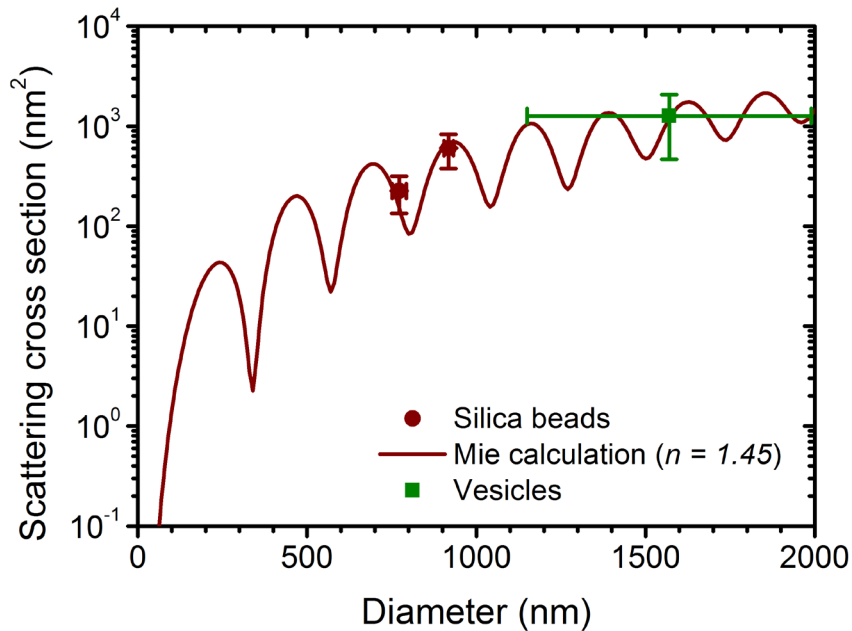
- BxPC-3 vesicles have different Raman spectra
- composition of vesicles from one cell type differs

# Results – Single vesicle trapping comparison



- Differences obtained! However,
  - only 12 single vesicles were trapped
  - low signal-to-noise ratio
  - vesicle size is unknown

# Estimation of vesicle size by elastic scattering



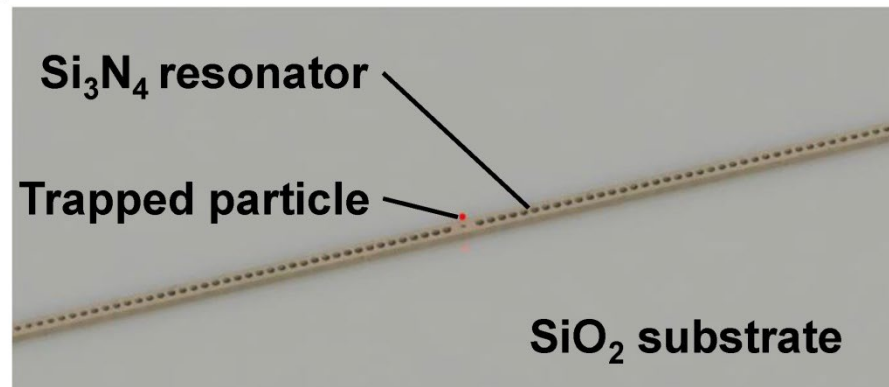
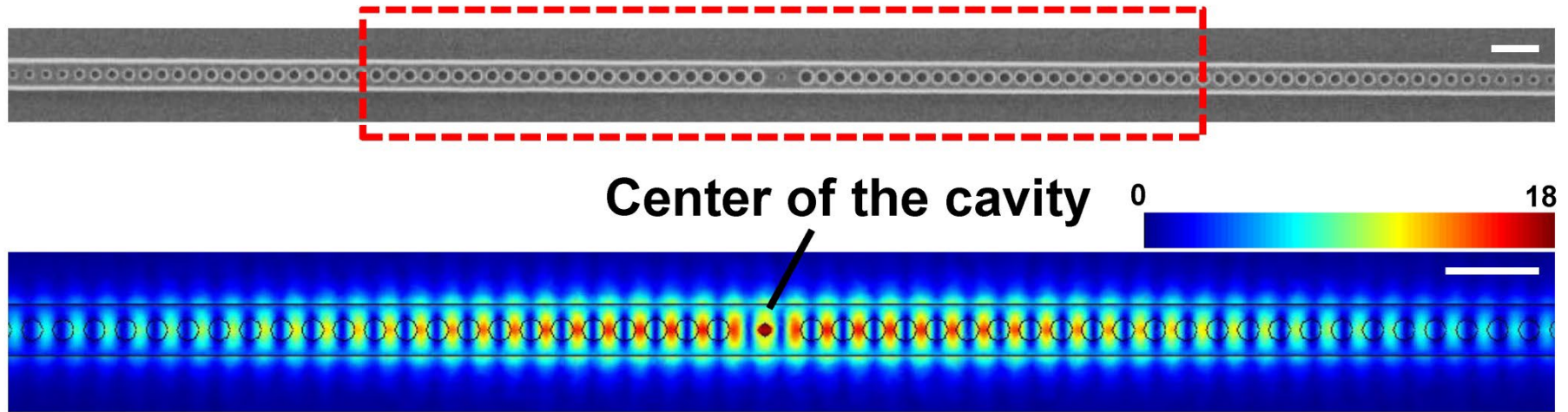
● vesicle diameter  $> 1 \mu\text{m}$



# Conclusion and discussion

- measured Raman spectrum of single vesicles
  - composition of vesicles from one cell type differs
  - more measurements on single vesicles required
- diameter of trapped vesicles  $> 1 \mu\text{m}$ 
  - trapping of smaller vesicles required

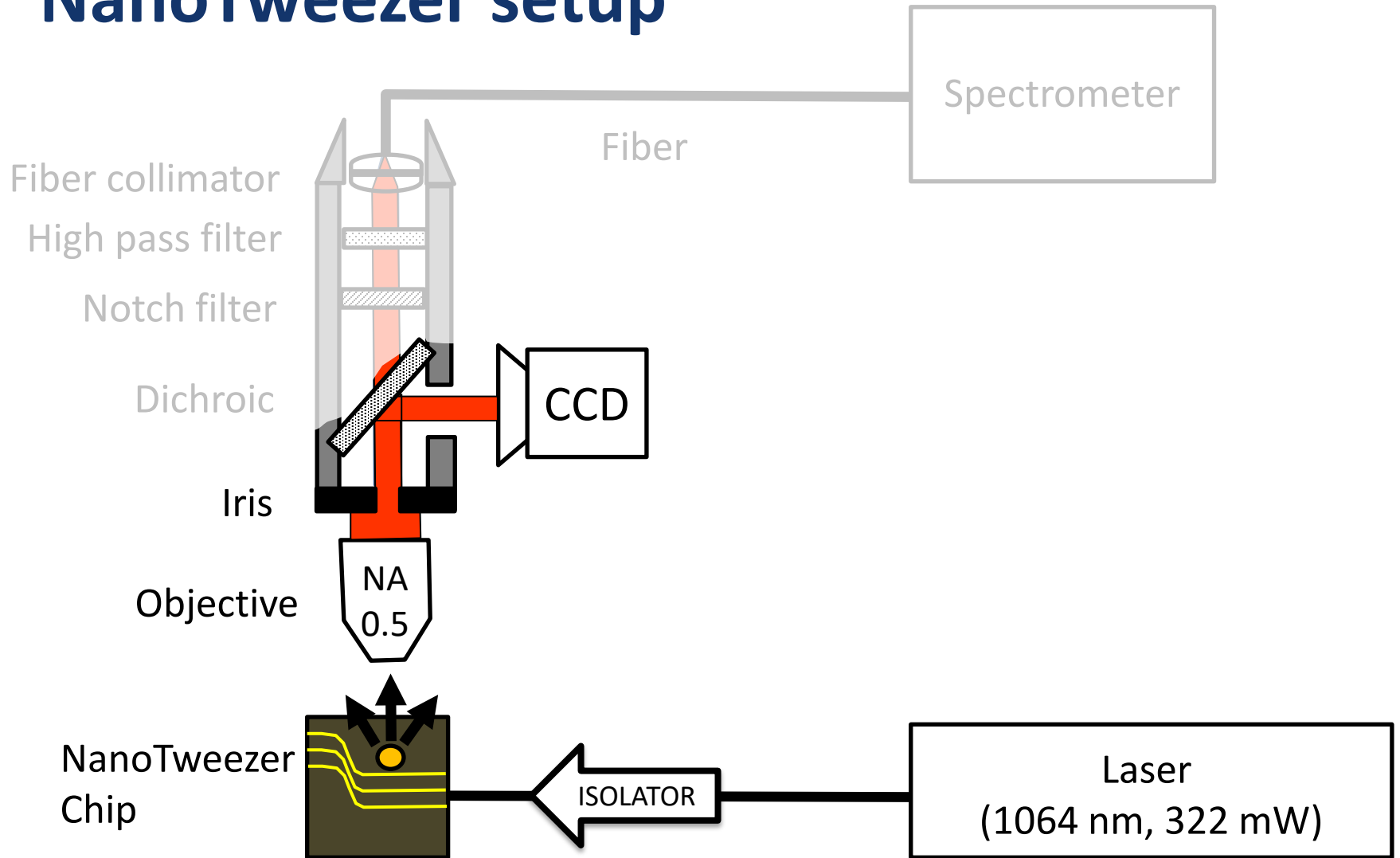
# Outlook: use NanoTweezer to trap vesicles



# Optofluidics NanoTweezer



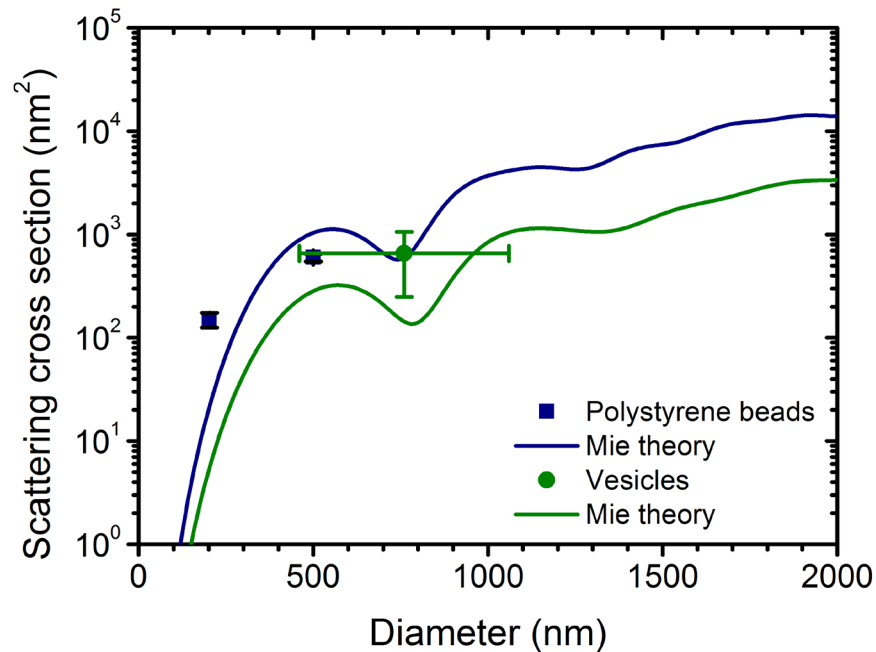
# NanoTweezer setup



# Trapping urinary vesicles with a NanoTweezer



# Size estimation of vesicles trapped by the NanoTweezer



# Outlook: Raman spectroscopy of single tumor vesicles with Nanotweezer

