# Flow cytometry standardization by size and refractive index determination

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#### **Conflicts of interest**

 Edwin van der Pol and Frank Coumans are cofounder and stakeholder of **EXOMETRY**

#### **Outline**

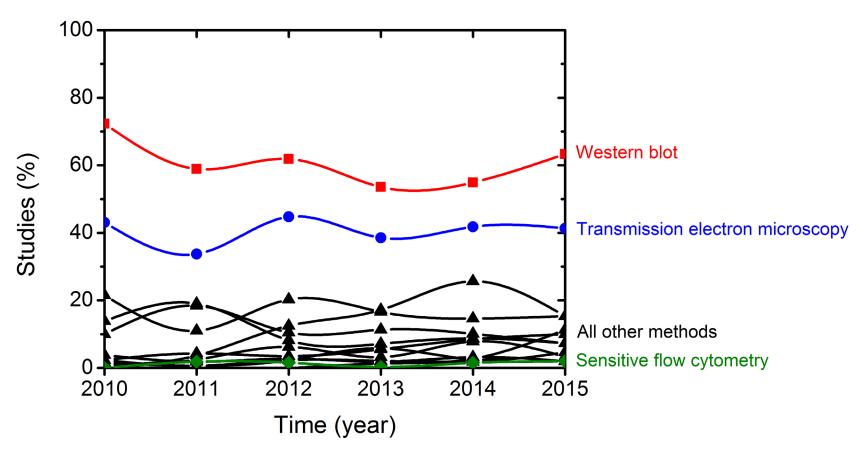
- standardization of flow cytometry measurements on extracellular vesicles
  - > motive
  - by size determination
  - > by size and refractive index determination
- Summary



# Extracellular vesicles (EV)

# How do we study EV?





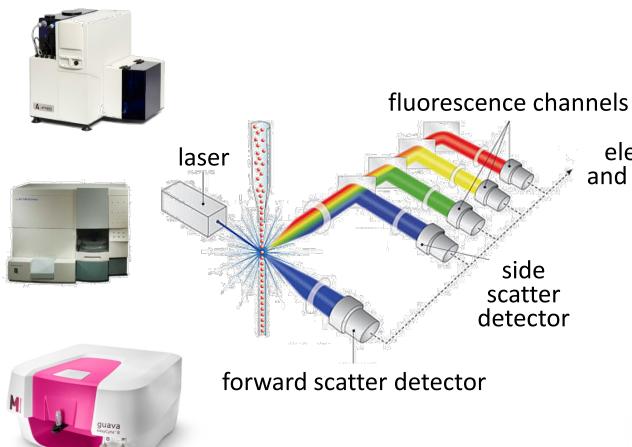
# Motivation to detect single EV

- EV are heterogeneous
- study the contribution of all EV, including rare EV



# Flow cytometry











#### Standardization is boring (biologists, clinicians)





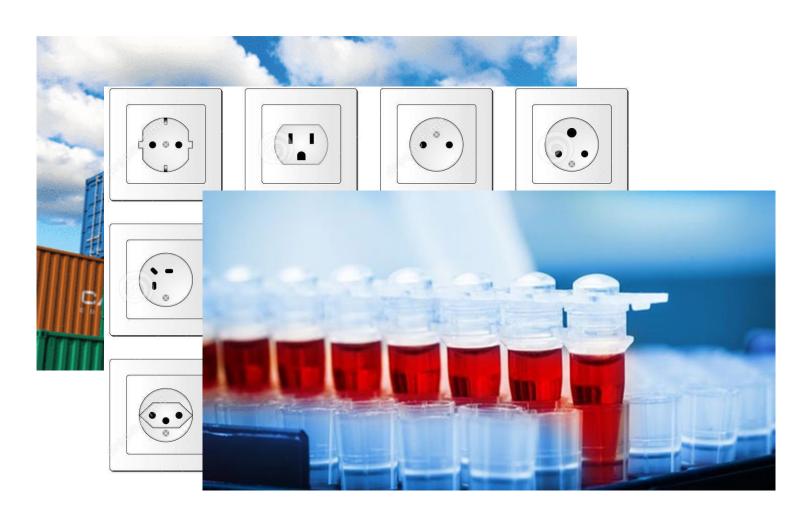
#### Standardisation is exciting (metrologists, physicists)

#### **BESSYII**

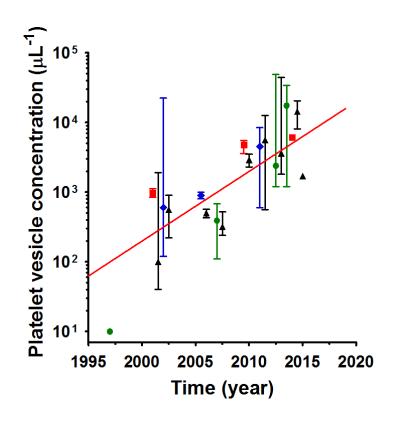


0.31 nm X-rays to size EV\*
(flow cytometers typically use 488 nm light)

# Standardization is important (everybody)



# Standardization is difficult (EV-field)



"Gąsecka's law"

- reported concentrations of plasma EV differ >10<sup>6</sup>-fold
- clinical data cannot be compared

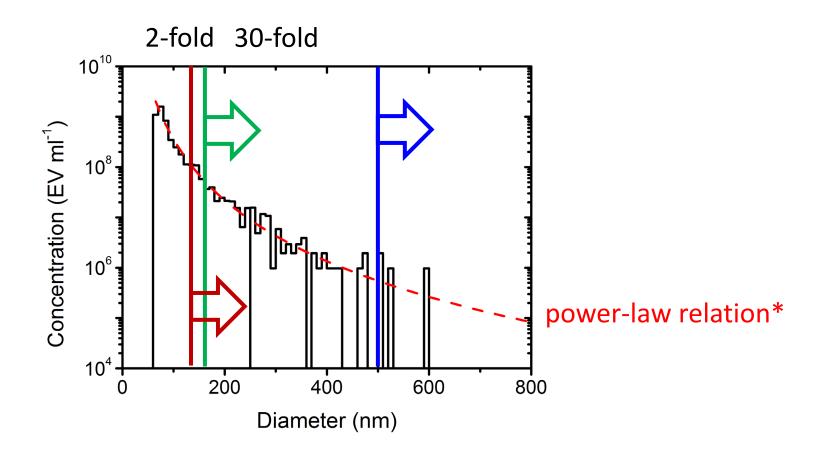
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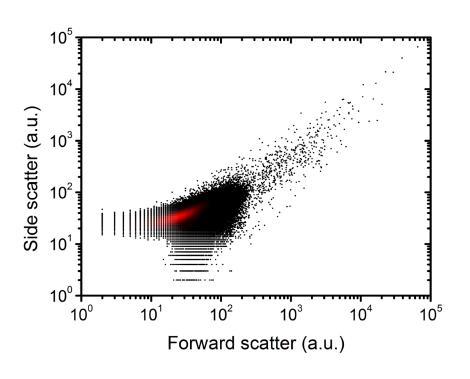


# **Problem 1: instruments differ in sensitivity**



# **Problem 2: arbitrary units**

#### same population of erythrocyte EV



Side scatter (a.u.)

Apogee A50-micro

**Becton Dickinson FACSCanto II** 

#### Goal

 obtain reproducible measurements of the EV concentration using different flow cytometers



#### Study comprises 33 sites (64 instruments) worldwide



# Approach scatter-based standardization

- measure EV reference sample and controls
- determine flow rate
- scatter (a.u.) → diameter (nm)
  - measure METVES-beads
  - Exometry software obtains scatter to diameter relation
  - Exometry software provides EV size gates
- apply EV size gate to software (e.g. FlowJo) and report concentrations

# **EV** reference sample

- erythrocyte EV from blood bank concentrate
  - ➤ CD235a-FITC labeled
  - > trigger on most sensitive scatter channel
  - > exclude EV similar to isotype

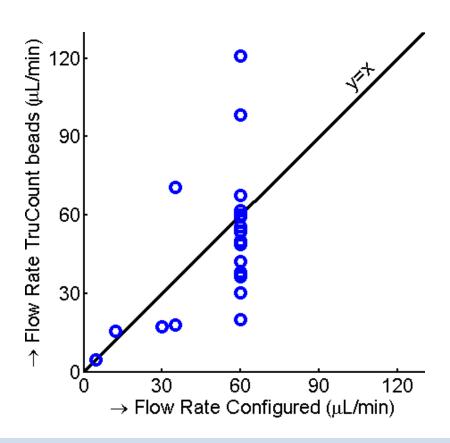


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#### **Determine flow rate**

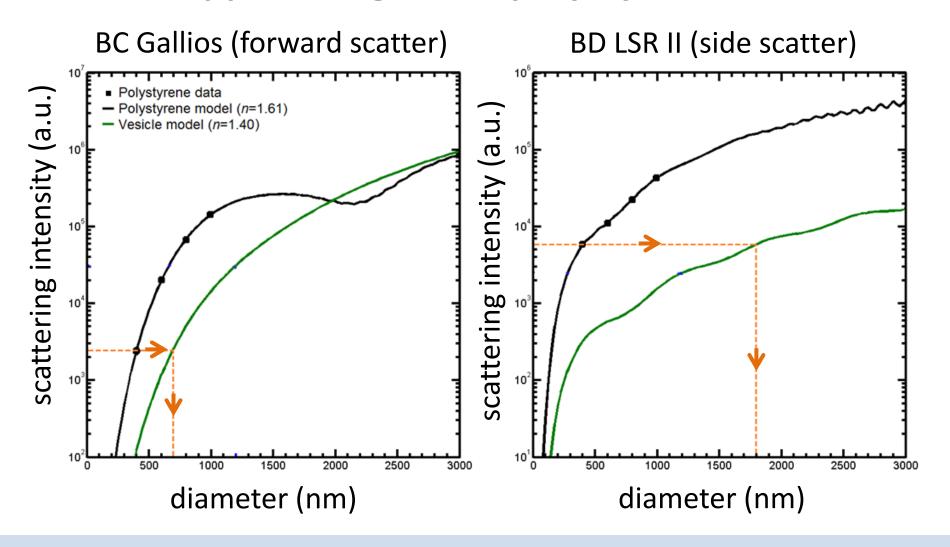
$$concentration = \frac{\text{\# of EV}}{\text{flow rate} \times \text{measurement time}}$$



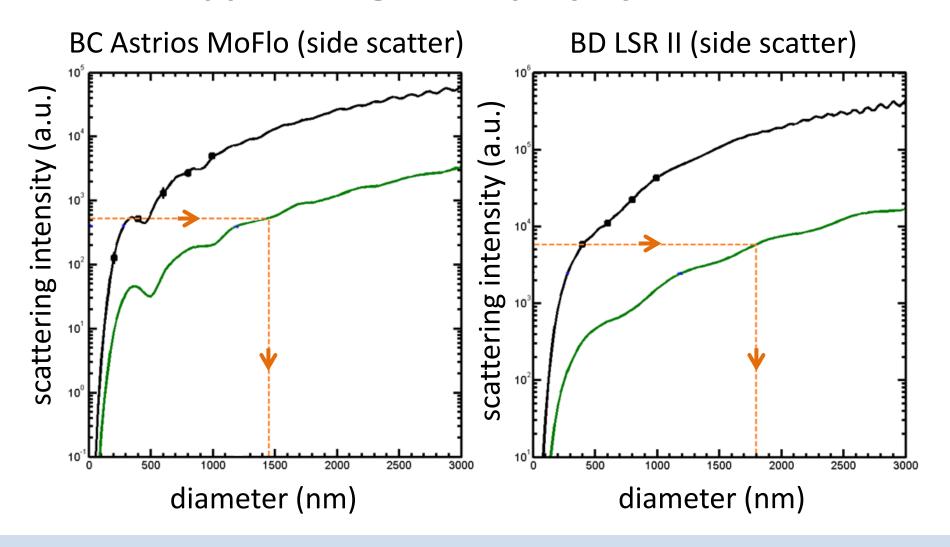
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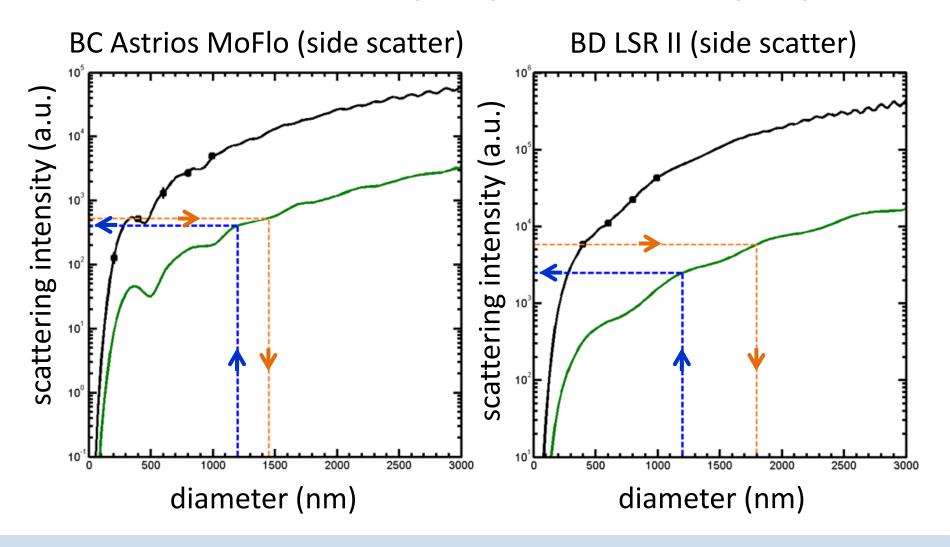
# Earlier approach: gate on polystyrene beads

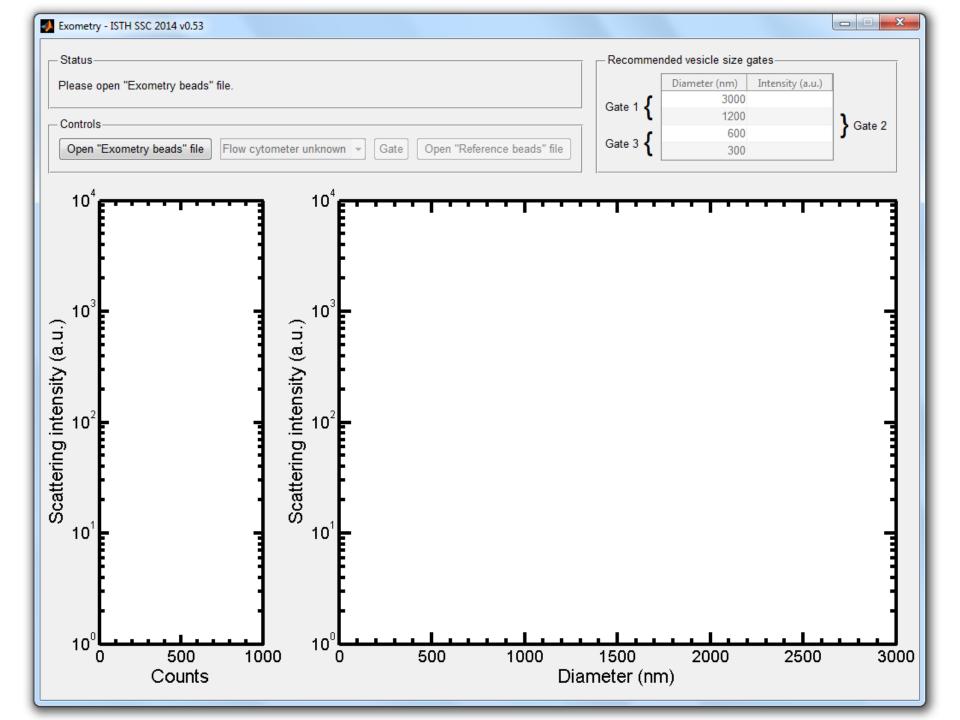


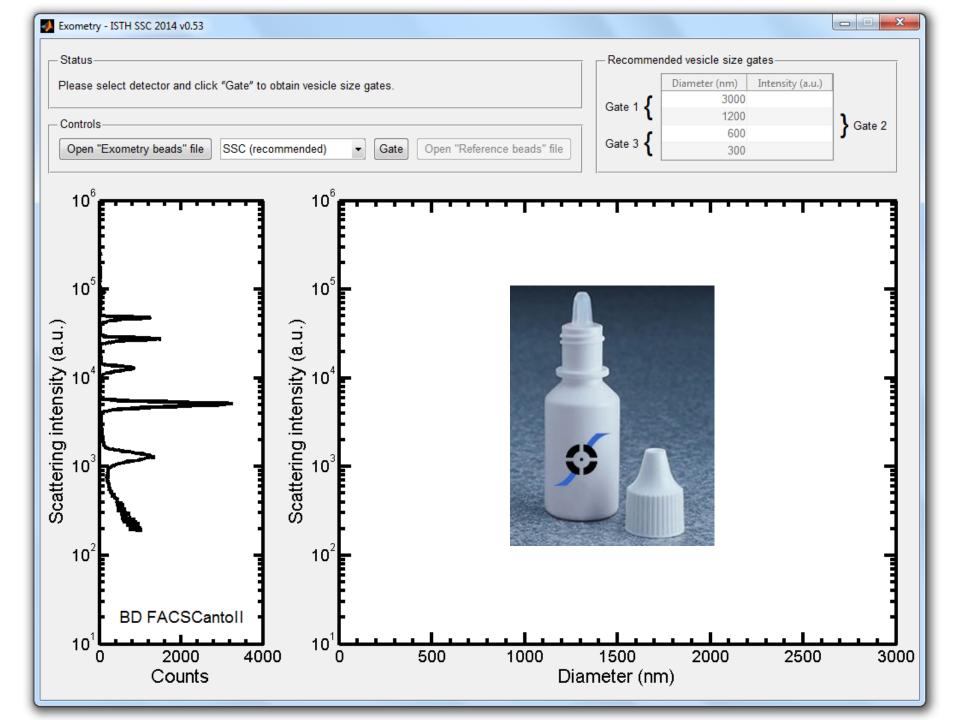
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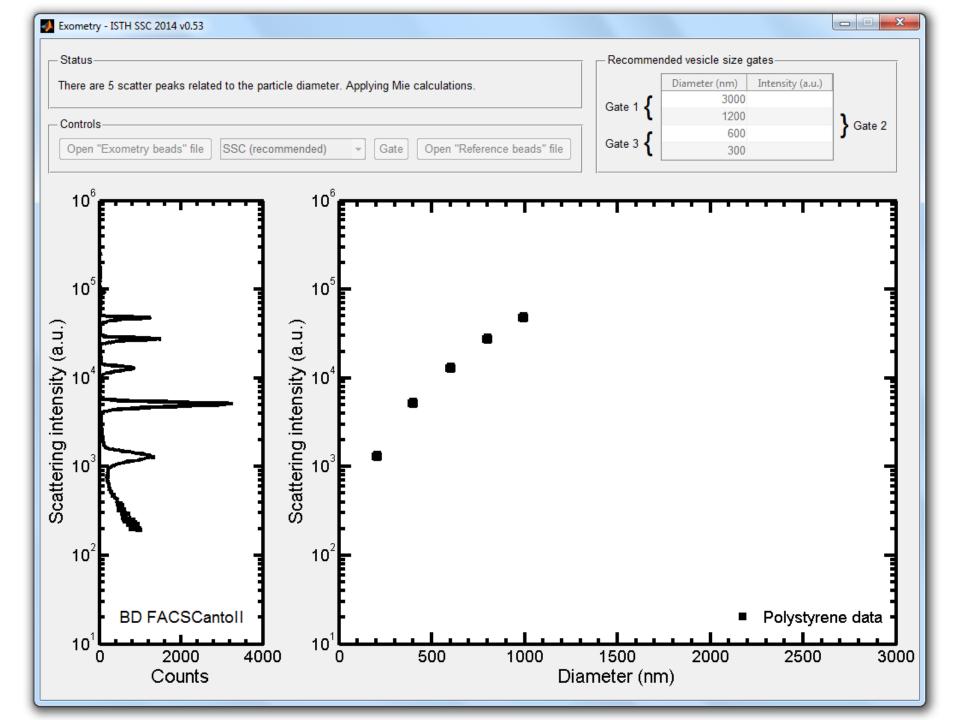


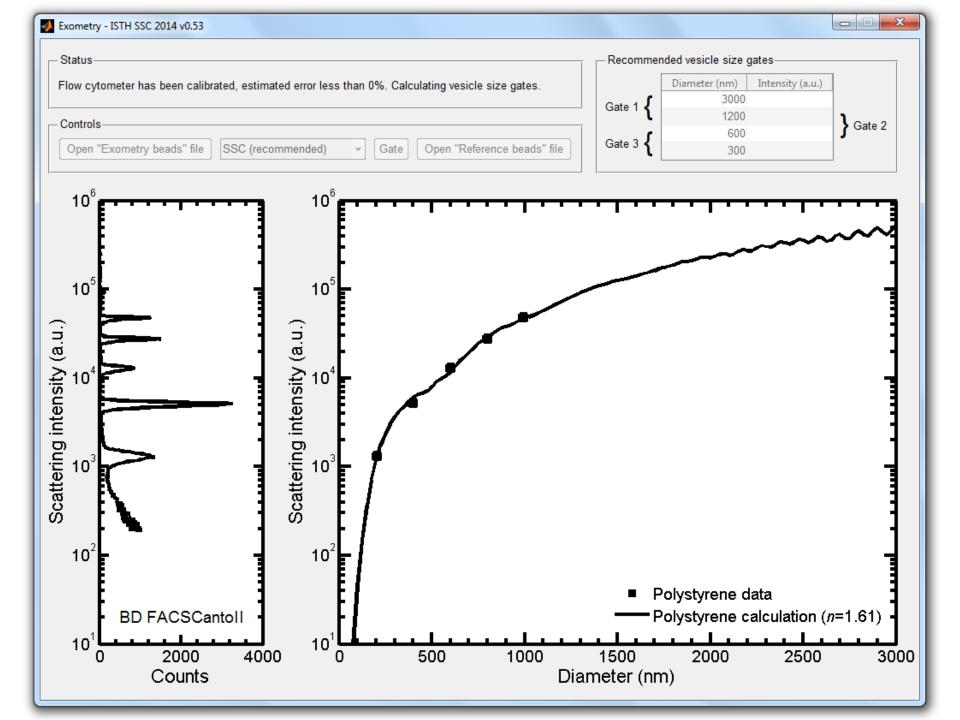
# 2016: relate scatter (a.u.) to diameter (nm)

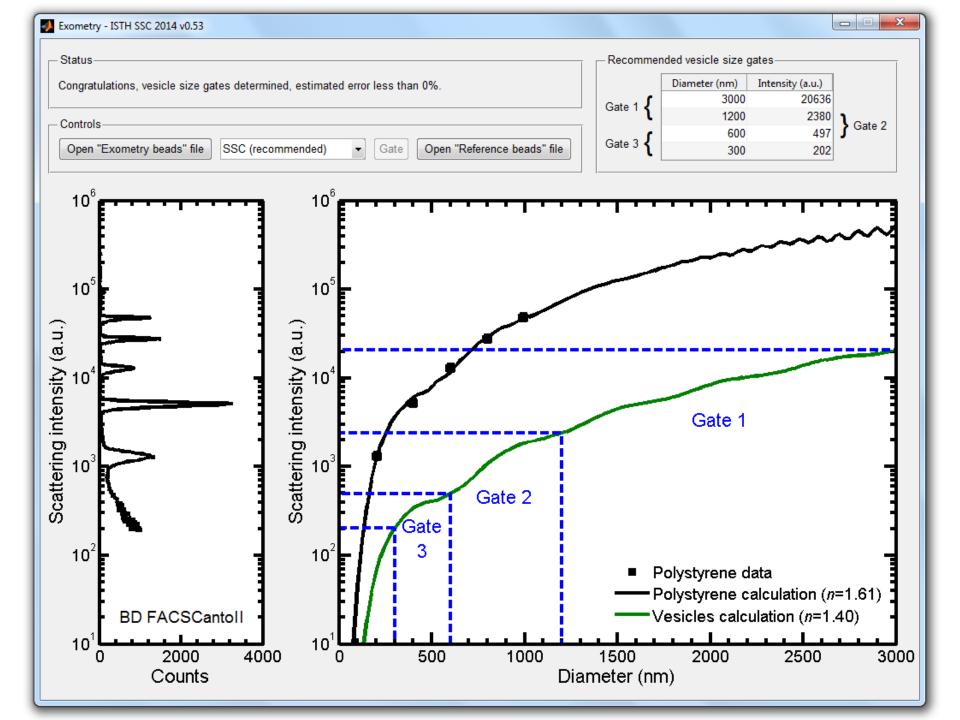


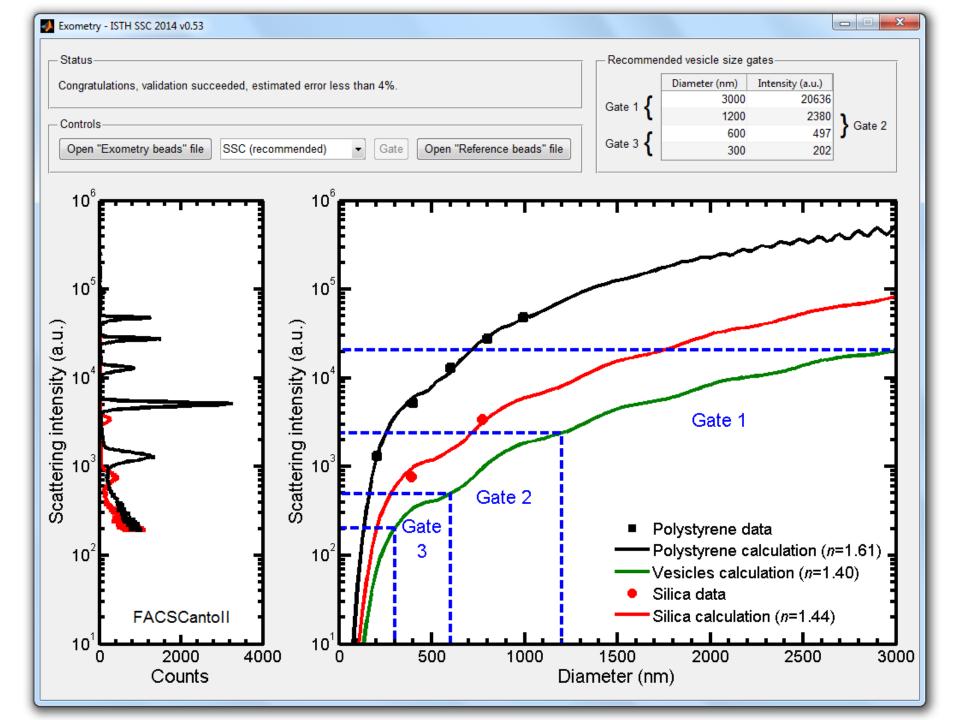


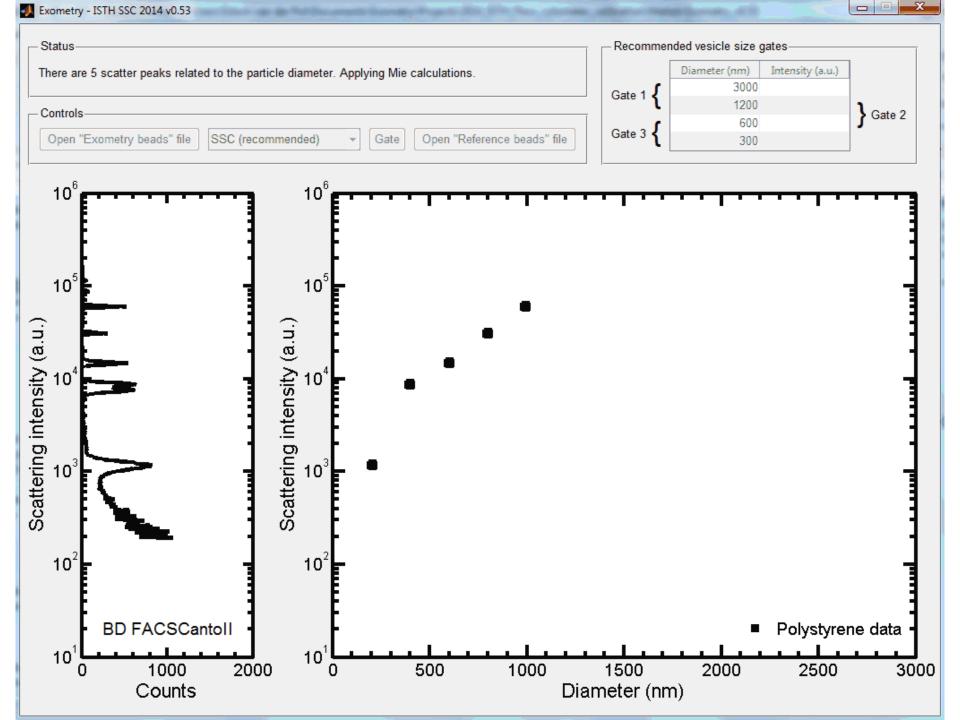




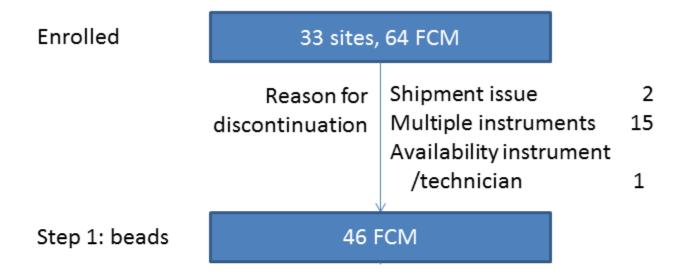






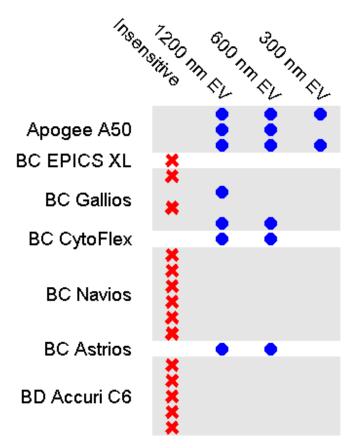


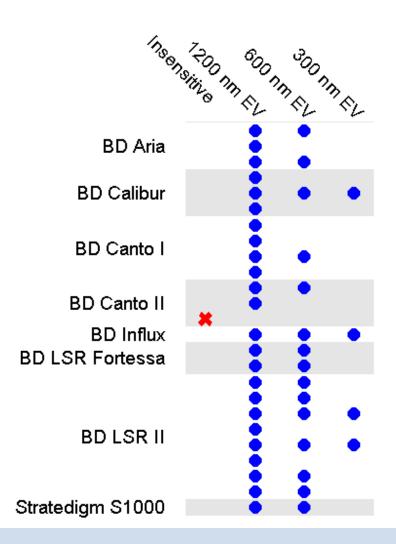
# **Exclusion of flow cytometers (FCM)**



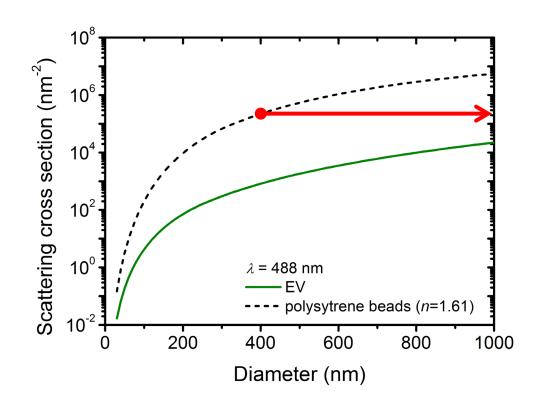
# Sensitivity of 46 flow cytometers in the field

= unable to detect 400 nm polystyrene beads



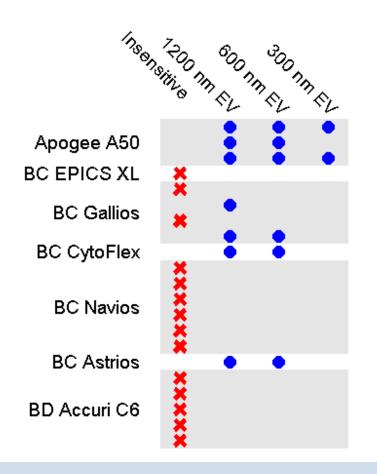


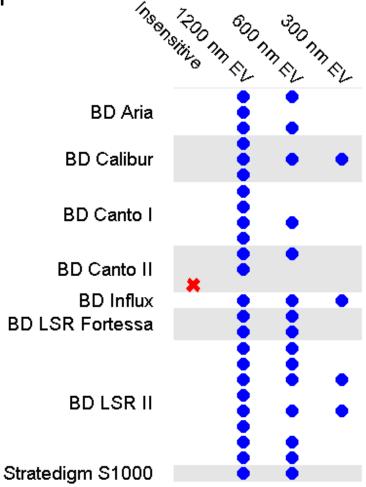
# 400 nm polystyrene beads scatter more than 1,000 nm EV



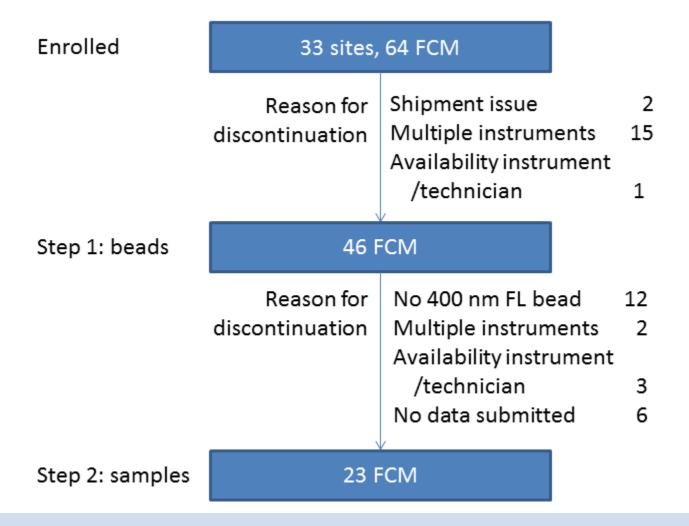
# Sensitivity of 46 flow cytometers in the field

■ = unable to detect EV < 1000 nm
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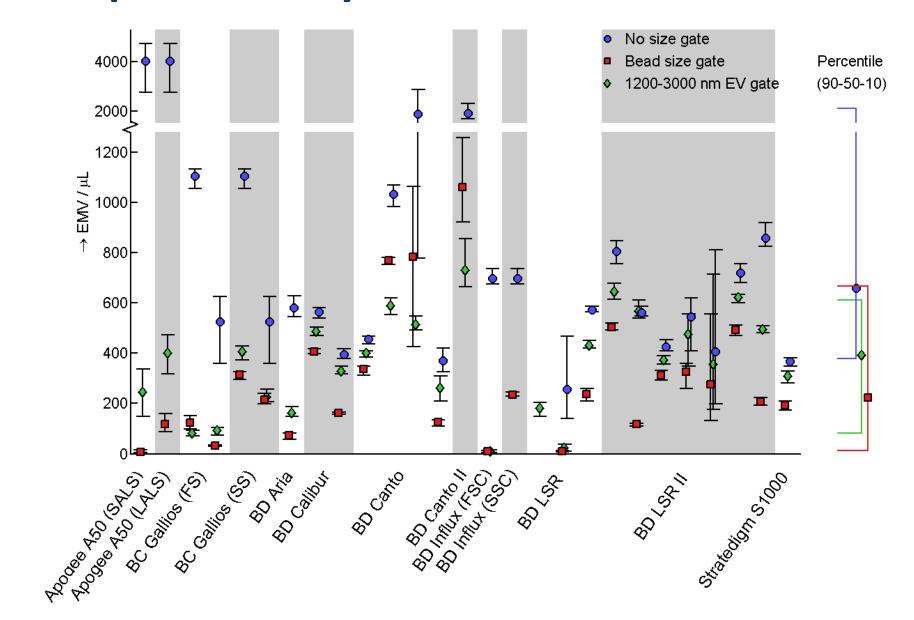
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# Reproducibility of 1200-3000 nm EV



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%CV	All	Side scatter	
		only	scatter only
Gate on beads	74%	60%	80%
Gate on EV size with light scatter theory	59%	42%	92%

# Conclusions standardization by sizing

- flow rate calibration is essential
- many flow cytometers used in EV research do not detect EV by scatter-based triggering
- EV size gate by Mie theory (CV=59%) leads to better reproducibility than gate on beads (CV=74%)

# Discussion standardization by sizing

- assumption of EV size gate by Mie theory
  - > EV have similar refractive index of 1.4
- discrepancy between FSC and SSC
  - due to incorrectly selected refractive index?
- standardization of EV sizes <1200 nm ineffective</li>

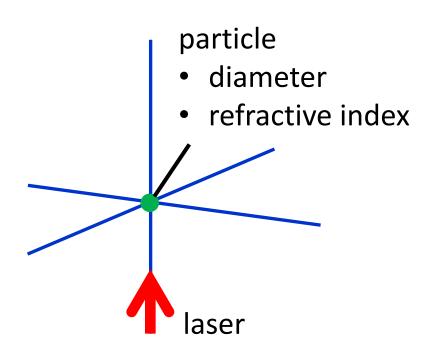
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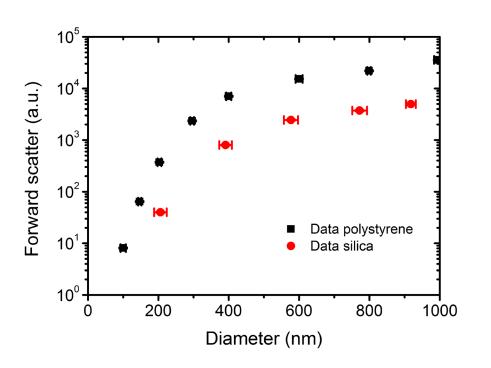
#### Size and refractive index determination of EV

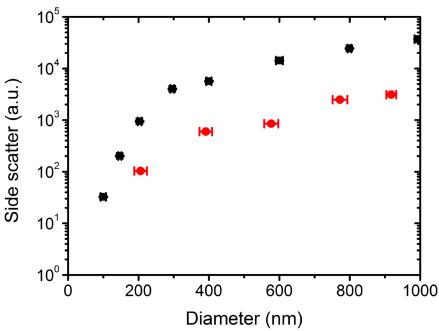


# **Approach**

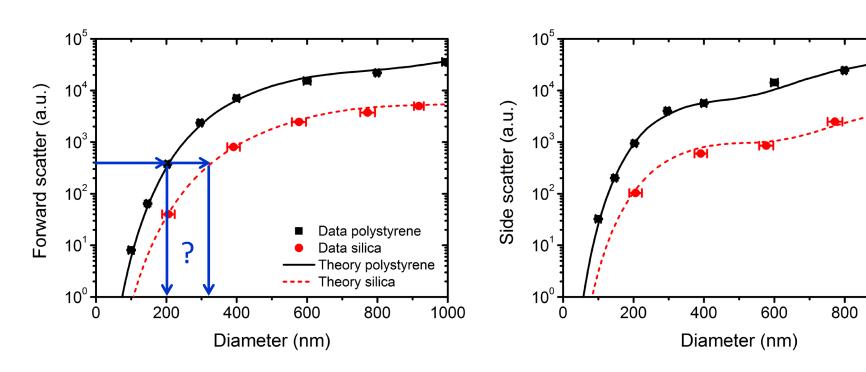
- calibrate instrument (Apogee A50-micro)
  - calibrate FSC and SSC
  - derive size from Flow Scatter Ratio (Flow-SR = SSC/FSC)
  - derive refractive index from size and FSC
- validate Flow-SR
  - beads mixture
  - > oil emulsion
- apply Flow-SR
  - > EV and lipoprotein particles from blood

### Calibrate forward scatter and side scatter





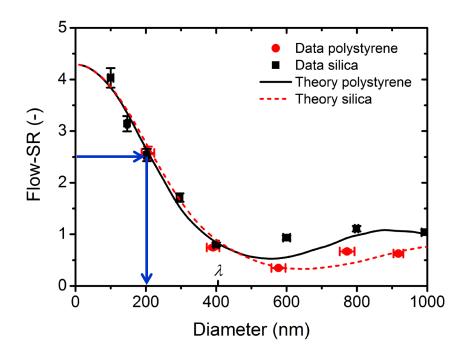
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Flow-SR = 
$$\frac{\text{side scatter}}{\text{forward scatter}}$$

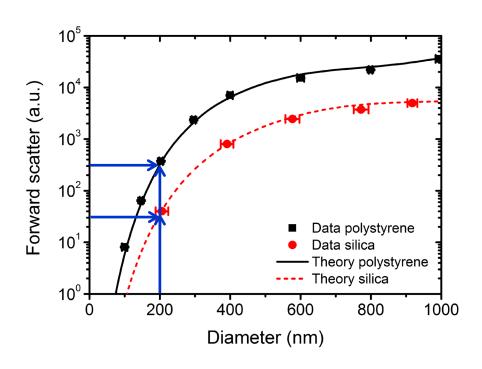
1000

#### **Derive size from Flow-SR**



Flow-SR = 
$$\frac{\text{side scatter}}{\text{forward scatter}}$$

#### Derive refractive index from size and FSC

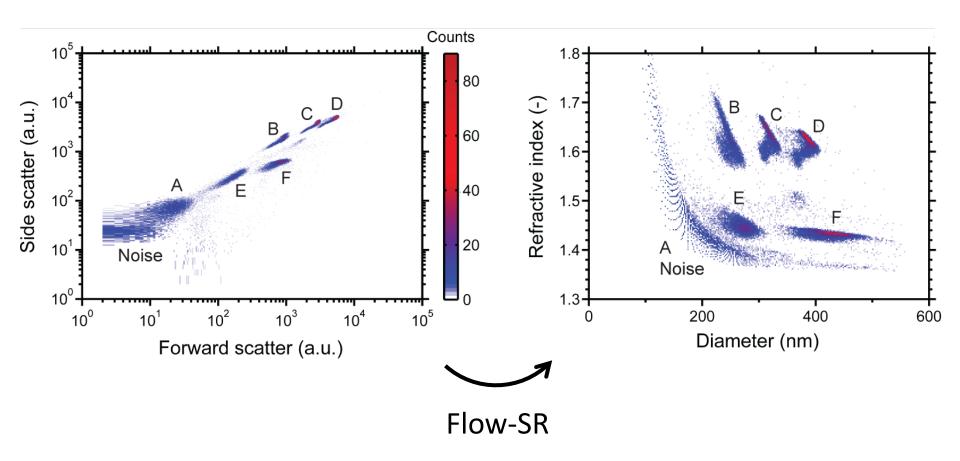


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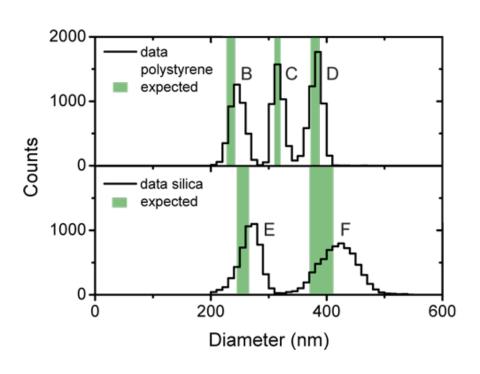
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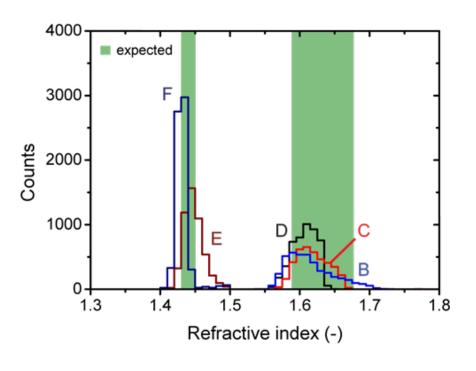
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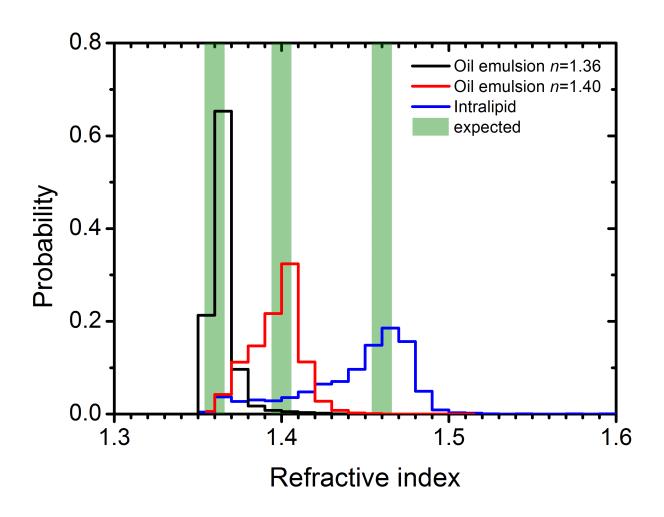




measurement error < 8% CV < 8%

CV < 2%

### Validate Flow-SR with oil emulsions

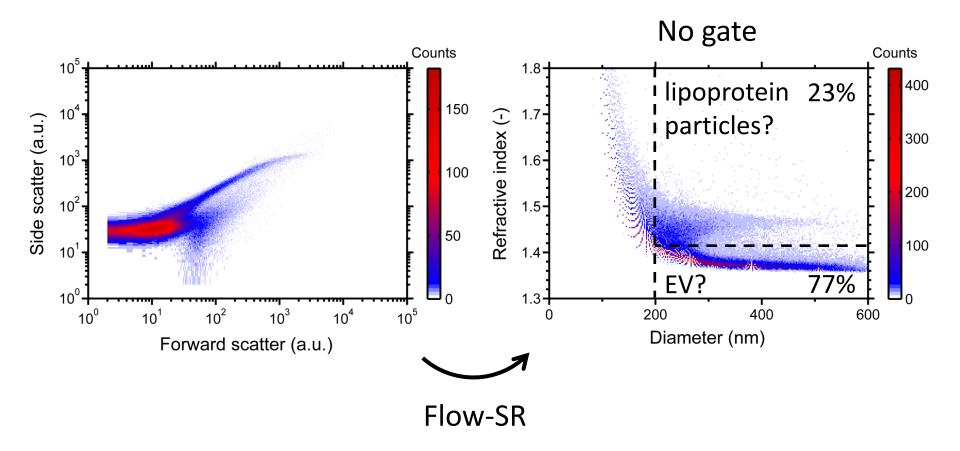


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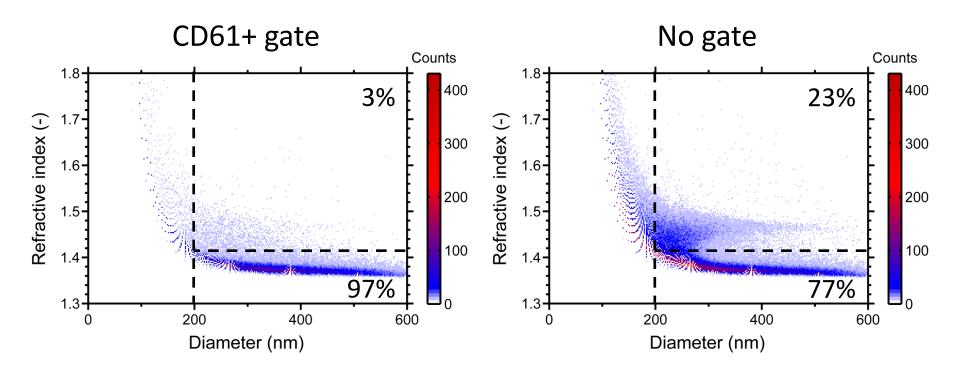
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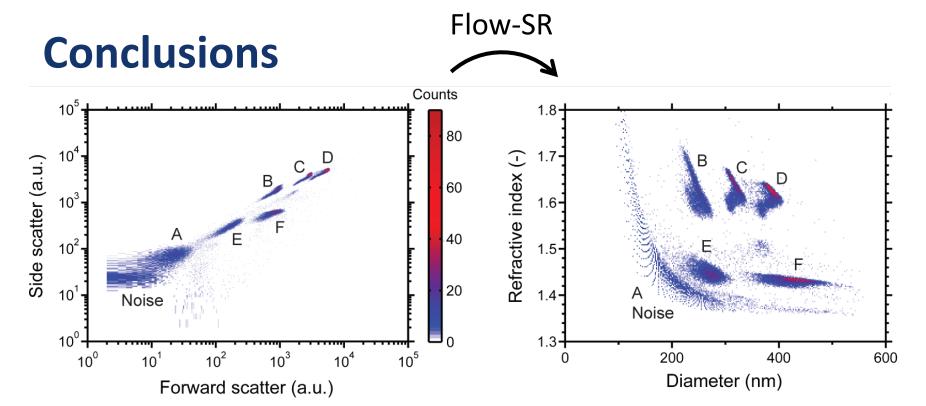
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### Supernatant of outdated platelet concentrate



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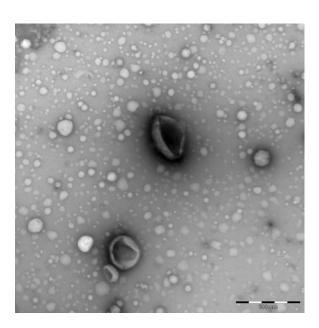




- Flow-SR enables size and refractive index determination of nanoparticles by flow cytometry
  - data interpretation and comparison
  - label-free identification

# **Summary**

- extracellular vesicles contain clinical information
- standardization is essential
- scatter provides valuable information
  - > size
  - > refractive index



# Acknowledgements

- Vesicle Observation Center
   Academic Medical Center
   University of Amsterdam
- Laboratory Experimental Cancer Research, Ghent University
  - Olivier de Wever
  - An Hendrix
- Software and beads by <u>exometry.com</u>
- More info: <u>edwinvanderpol.com</u>













